



Synthesis and Characterization of Ammonium Benzilate Bioactive Ionic Liquids and Their Antimicrobial Activity

Md. Ismail Hossain¹ and Ajoy Kumer^{1*}

¹*Ionic Liquid Research Laboratory (ILRL), Department of Chemistry, University of Chittagong, Chittagong 4331, Bangladesh.*

Authors' contributions

This work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.

Article Information

DOI: 10.9734/AJOPACS/2017/38386

Editor(s):

(1) Victor G. Zavodinsky, Professor, Department of Physics, Institute of Materials Science, Russia.

Reviewers:

- (1) Simone Carradori, "G. d'Annunzio" University of Chieti-Pescara, Italy.
(2) Joseph Sloop, Georgia Gwinnett College, USA.
(3) Fatma Kandemirli, Kastamonu University, Turkey.
(4) Claudia Araceli Contreras Celedón, Universidad Michoacana de San Nicolás de Hidalgo, México.
(5) Vinit Raj, Babasaheb Bhimrao Ambedkar University Lucknow, India.

Complete Peer review History: <http://www.sciencedomain.org/review-history/22953>

Original Research Article

Received 24th October 2017
Accepted 17th January 2018
Published 31st January 2018

ABSTRACT

The ammonium benzilate Ionic Liquids (ILs) was synthesized by the neutralization reaction of different cations of ethanolamine, dimethylethanolamine, and triethanolamine with benzoic acid. The reaction is monitored by thin layer Chromatography. The synthesized compounds were purified. Then the synthesized ILs was characterization by FT-IR Spectroscopy and ¹H Nuclear Magnetic Resonance (NMR) Spectroscopy. Antibacterial screening of the test ILs was carried out with six human pathogenic bacteria, such as *Bacillus cereus*, *Staphylococcus aureus*, *Escherichia coli*, *Salmonella typhi*, *Pseudomonas aeruginosa* and *Shigella dysenteriae* and two phytopathogenic fungi like *Aspergillus niger* and *Rhizopus azzahra*. The minimum inhibitory concentration (MIC) was determined to establish a good bioactive molecule.

Keywords: FT-IR; NMR and antimicrobial activity.

*Corresponding author: E-mail: kumarajoy.cu@gmail.com;

1. INTRODUCTION

Ionic Liquids (ILs) was introduced almost a century ago by the observation of Paul Walden in a neutralisation reaction of ethanolanmonium nitrate [1]. ILs are organic salts mainly composed of organic cations and inorganic anions, which are per definition liquid below 100°C and exhibit in most cases relatively low viscosities [2]. The definition allows distinguishing them from a classical molten salt, which is mostly high-melting, highly viscous, highly polar, very low vapour pressures, thermally stable, wide kinetic control high thermal conductivity, a large electrochemical window and very corrosive substance [3-5]. Conventionally, ILs typically contains bulky asymmetric organic cations, such as imidazonium, pyridinium, pyrrolidinium, quaternary ammonium or tetraalkylphosphonium with very low symmetry to lead weak intermolecular interactions and low charge densities [6-7]. Quaternary ammonium based ILs is the most expected properties of ILs at which organic amine base are used as cation sources to produce salts with different types of carboxylic acids or inorganic acid as anion sources. Much quaternary ammonium ILs could be prepared to "tune" the physical properties such as high melting point, water miscibility, conductivity and viscosity by selection and modification of the anion and cation combination of the IL. ILs have attracted much attention due to the widespread potential for practical applications such as heat transfer and storage medium in solar thermal energy systems as well as for many areas such as fuel cells, rechargeable batteries and "green solvents" in the chemical process [8].

In the race to synthesize new pharmaceutical drugs, ILs has attracted a great deal of attention due to their variety of potential pharmaceutical applications [8-9]. The strategy of ILs can take advantage of the dual nature (by controlling the alkyl length of cations [10,11,12]. Quaternary ammonium salts (quats) are an economically advantageous class of industrial compounds [13]. They have surface-active properties (discrete ions) to realize enhancements which may include controlled solubility (e.g., both hydrophilic and hydrophobic) and are possible bioactivity [14]. The antimicrobial activity and toxicity towards both prokaryotic and eukaryotic micro-organisms are designed and changed [15]. ILs possesses anti-microbial activity known to be bioactive in nature [16-17]. The low melting point of ammonium salts was relatively rare in the

literature of antimicrobial activity during the last decades. The encouraging results of preliminary toxicological studies of synthesized ILs provide good opportunities for the development of biomedical applications for drug discovery [18]. The third generation ILs is active pharmaceutical ingredients [19]. The synthesized third generation ILs provides good opportunities for the replacement of natural bioactive molecule acted as the synthetic bioactive molecule with the satisfactory of green chemistry [20-21].

2. EXPERIMENT

2.1 Materials and Reagents

All the chemicals were of research grade and used without further purification unless otherwise stated. All the solvents were obtained upon distillation before use. Ethanolanine (Merck KGaA), Dimethylethanolanine (Merck KGaA), Triethanolamine (MarckKGaA) and Benzilic acid (Merck KGaA) were analytical grade commercial products. Thin Layer chromatography powder (Merck KGaA), standard antibiotic Gentamycin were purchased for the reaction workup. FT-IR spectrophotometer, (SHIMADZU, Japan, range 600 -4500 cm⁻¹) was used with KBr disc technique. The synthesis and the characterization were done at the Department of chemistry at University of Chittagong, and Chittagong-4331, Bangladesh. The antimicrobial activity was at the Department of Microbiology in University of Chittagong, and Chittagong-4331, Bangladesh. The ¹H NMR Spectroscopy was done at Iwate University, Japan.

2.2 Ionic Liquids Synthesis and Purification

The synthesis of the alkanol or alkyl ammonium ILs consists in an acid-basic neutralization reaction [22]. The base, in this case, ethanolamine, dimethylethanolanine, and triethanolamine were added under stirring in a slow dropwise about 20-25 minute maintaining the temperature using ice bath from exothermic heat release on a glass flask with benzilic acid. Then the mixture was stirred for (24-28) hours at room temperature, to obtain a viscous clear liquid. The reaction was monitored by Thin layer chromatography (TLC).The reaction products are an ester and a salt of ammonium [13, 22-23]. The purification process consists of a strong

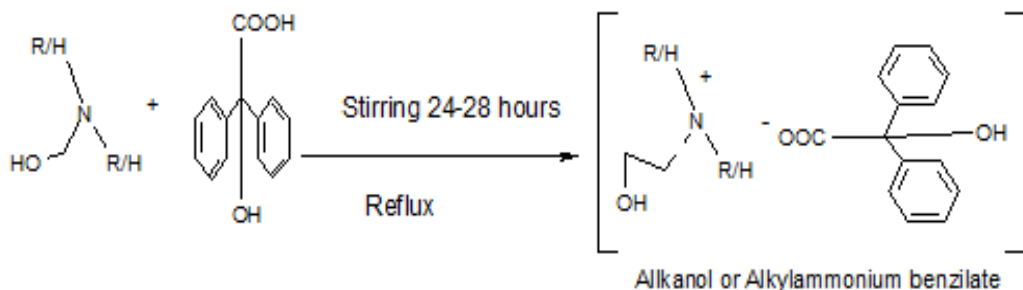


Fig. 1. General reaction scheme for synthesis of ILs

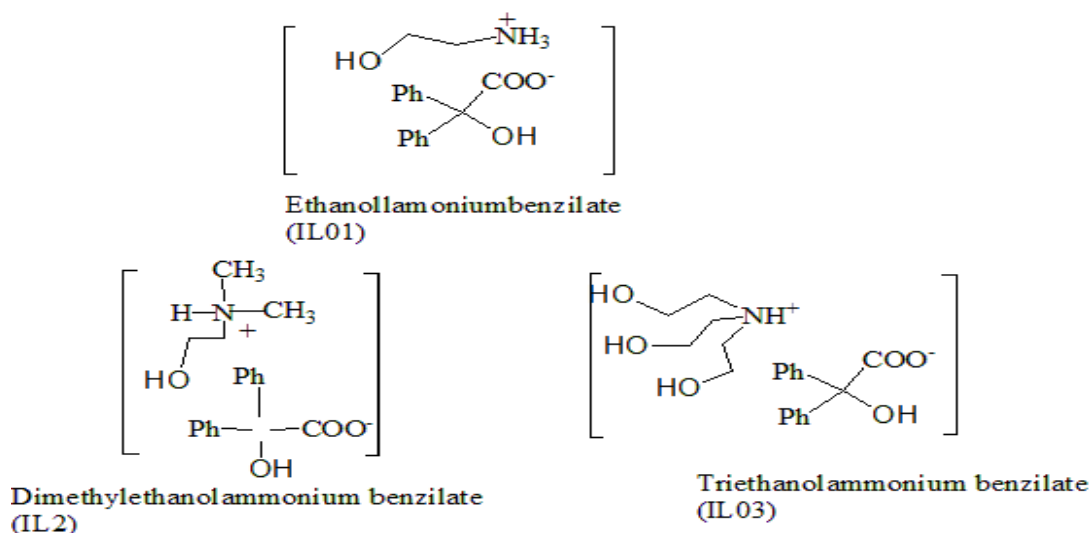


Fig. 2. Synthesized ammonium based IL

agitation and slight heating, at 323.15K, for the vaporization of impurities (residual non-reacted and water) under a vacuum of 20 kPa. Humidity below 0.1% was obtained after this purification process[22-23] and the liquids presented a limp and viscous appearance. The ammonium salt formation was proved in by FT-IR spectroscopy by using a Shimadzu IR and ^1H NMR spectroscopy. The structure of synthesized IL is shown in Fig. 2.

3. RESULTS AND DISCUSSION

3.1 FTIR Analysis

The presence of carboxylate (COO^-) peak is at $\sim 1597\text{ cm}^{-1}$ and $\sim 1338\text{ cm}^{-1}$ by N-H vibrations. The broad absorption around $\sim 3335\text{-}3441\text{ cm}^{-1}$ can be assigned the presence of OH groups. Then the C-O bond has also confirmed the peak

of $1260\text{-}1000\text{ cm}^{-1}$ in the carbonate salts and 2995 cm^{-1} is for the stretching of Ar-H bond. The presence of Ph-COO group has confirmed the peak at $\sim 1724\text{ cm}^{-1}$. The broad peak of ammonium in $3500\text{ to }2400\text{ cm}^{-1}$ confirms the formation of ammonium salt as ammonium based ILs.

Ethanolammonium benzilate (IL01), $[\text{C}_2\text{OHNH}_3]$ $[\text{Ph}_2\text{C}(\text{OH})\text{COO}]$, M.W.: 289.15, Yield (%): 91.0.

Found: %C = 66.40- 66.76, %H=6.62-6.98, % N= 4.78-5.03

Calculated: FT-IR (KBr) in cm^{-1} : 3414, 3001, 2947, 2885, 1620, 1558, 1377, 1068, 1018

$^1\text{H-NMR}$ chemical shifts: 2.0 (s, 1H, OH), 2.0 (s, 1H, OH), 3.52 (t, 2H, NCH_2), 4.27 (t, 2H, OCH_2), 7.0 (s, 3H, NH_3), 7.11-7.39 (m, 10H, 2Ph)

Dimethylethanolammoniumbenzilate (IL02),
 $[(CH_3)_2(C_2OH)NH_2] [Ph_2C(OH)COO]$, M.W. 318.19, Yield (%); 93.0.

Found: %C = 67.88- 67.93, %H=7.60- 7.46, % N= 4.40- 4.34

Calculated: FT-IR (KBr) in cm^{-1} : 3441, 3012, 2881, 1809, 1485, 1300, 1053, 752, 698

¹H-NMR chemical shifts: 2.0 (s, 1H, OH), 2.0 (s, 1H, OH), 2.80 (s, 6H, 2 \times NCH₃), 3.14 (t, 2H, NCH₂), 3.66 (t, 2H, OCH₂), 7.0 (bs, 2H, NH₂), 7.27-7.30 (m, 10H, 2Ph)

Triethanolammoniumbenzilate (IL03),
 $[(C_2OH)_3NH] [Ph_2C(OH)COO]$, M.W. 377.0, Yield (%); 68.0.

Found: %C = 63.62-63.23, %H=7.21-7.41, % N= 3.71-3.99

Calculated: FT-IR (KBr) in cm^{-1} : 3356, 2970, 2931, 2129, 1562, 1485, 1404, 1292, 1076.

¹H-NMR chemical shifts: 2.0 (s, 3H, 3 \times OH), 2.00 (s, 1H, OH), 3.43 (t, 6H, 3 \times NCH₂), 3.77 (t, 6H, 3 \times OCH₂), 7.10 (s, 1H, NH), 7.19-7.24 (m, 10H, 2Ph).

4. EXPERIMENT WITH ANTIMICROBIAL ACTIVITY

4.1 Preparation of IL Solutions in Different Concentrations

The required amount of the sample was measured in Digital balance with highly carefully so that no impurities were obtained. At first 20% solution was prepared to investigate the antimicrobial activity. Then by serial dilution process, 1000 mili-mole per Litre (mM/L), 750 mM/L, 500 mM/L, 250 mM/L and 100 mM/L solution was prepared and used to determine the MIC.

4.2 Antimicrobial Assay

A preliminary investigation of the antibacterial activities of pure ILs was performed through measurements of primary screening both the gram-positive and gram-negative bacteria. Antimicrobial screening of the 20 % solution of ILs was carried out with six bacterial pathogens, such as *Bacillus cereus*, *Staphylococcus aureus*, *Escherichia coli*, *Salmonella typhi*, *Pseudomonas aeruginosa* and *Shigella dysenteriae*. This method was carried

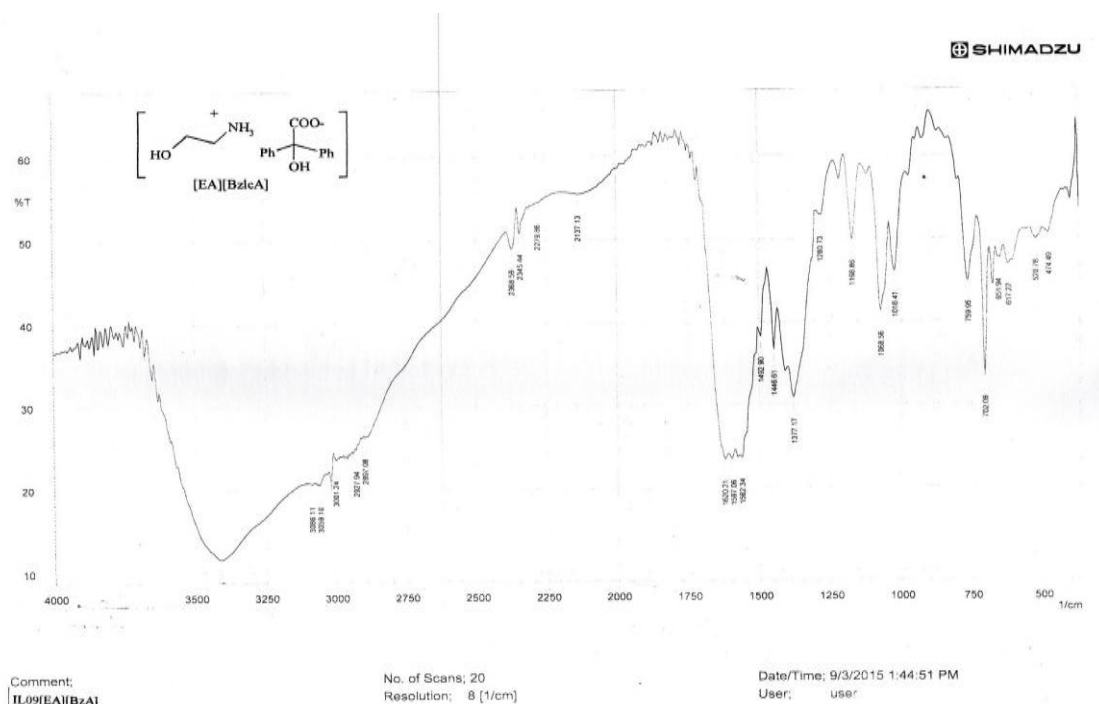


Fig. 3. FTIR peak of Ethanolammonium benzilate

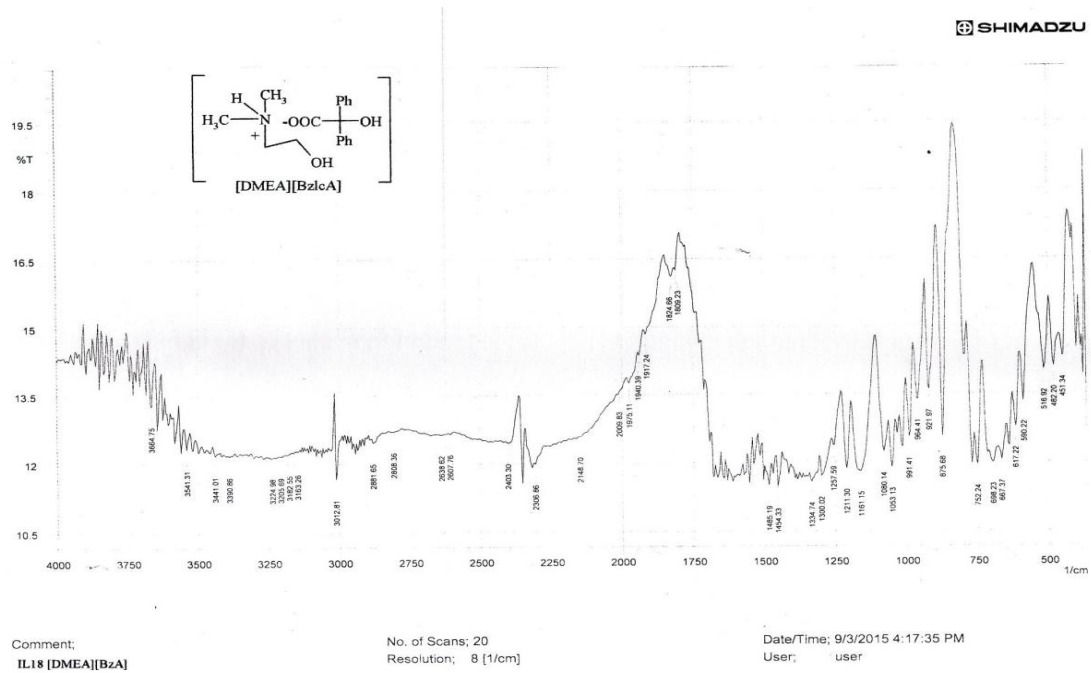


Fig. 4.

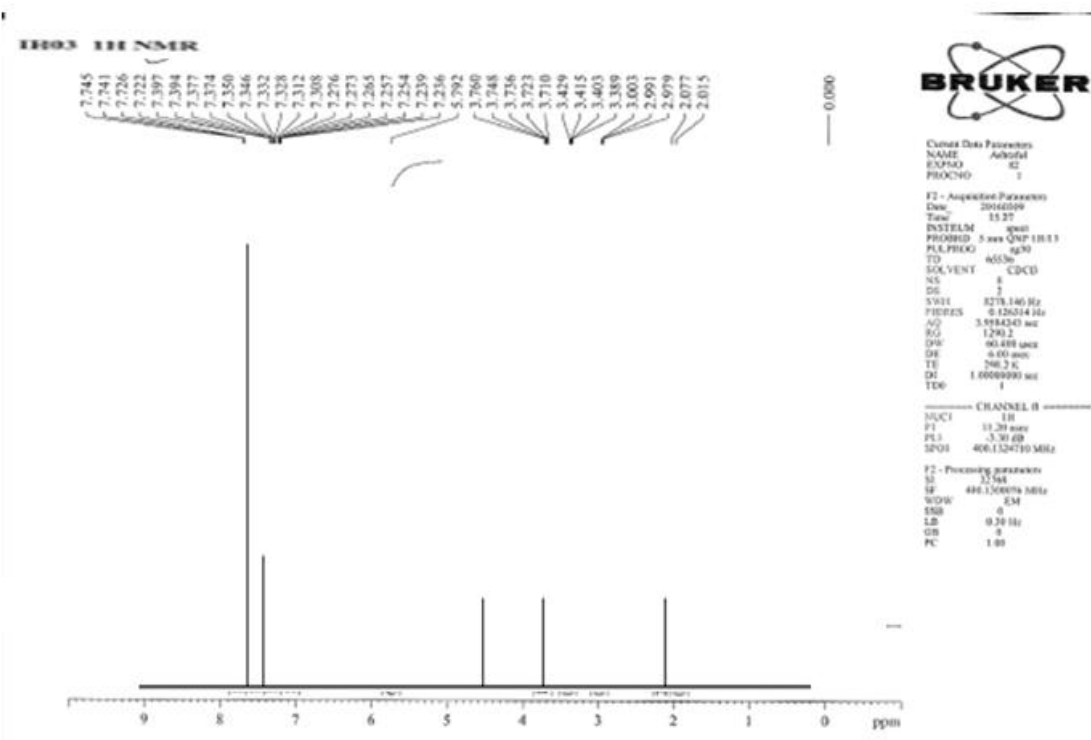


Fig. 5. 1H NMR for IL01

on via well diffusion method [24-25]. The diameter 5.0 mm that is the diameter of cork bacterial inhibition zone (subtracting the well borer) was measured in mm scale with

consideration ± 1.0 with all taking value. All the measurements were done in triplicate and the averages were listed in Table 1. The concentration for MIC determination was maintained for all ILs in 1000 mM/L, 750 mM/L, 500 mM/L, 250 mM/L, 125 mM/L and 75mM/Lin

distilled methanol through serial dilution technique. A control plate is always observed for the ILs if there is any significant inhibition occurred for the solvent. The results showed that all compounds had antimicrobial activity against bacterial pathogens used in this study.

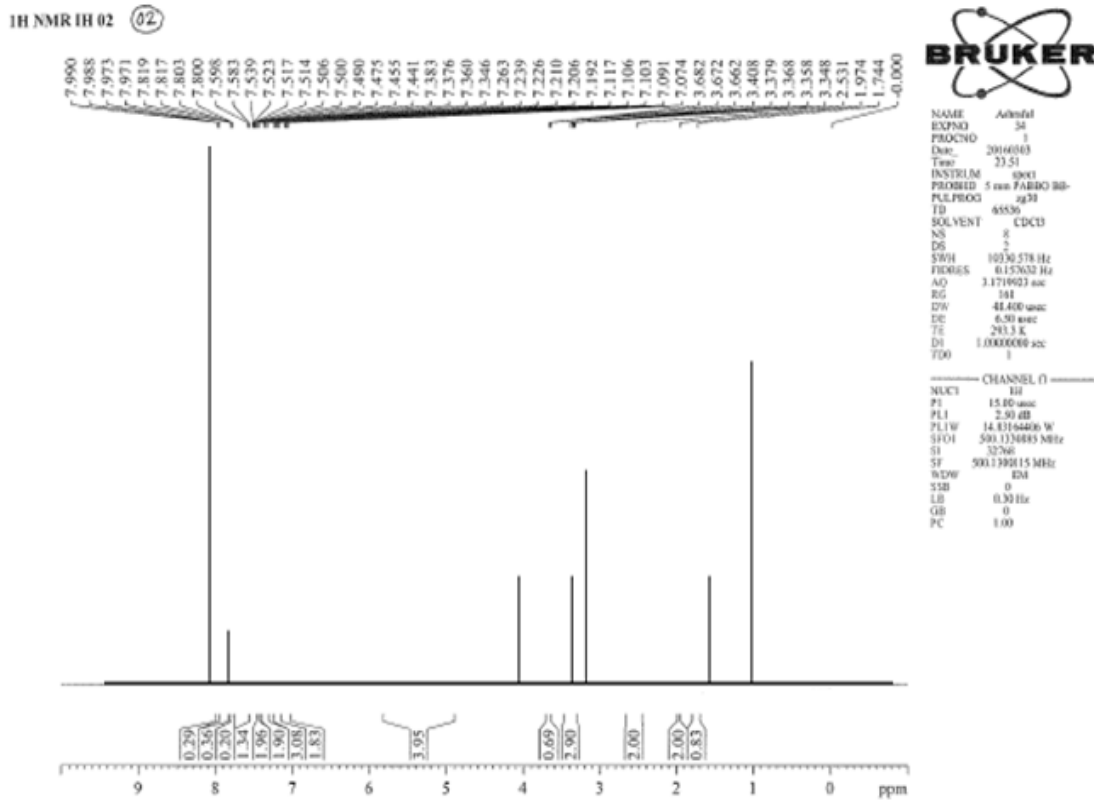


Fig. 6. 1HNMR for IL02

Table 1. Zone of inhabitation for 20% solution of ILs

List of pathogens and ILs	List of pathogens and ILs					
	<i>B. cereus</i>	<i>S. aureus</i>	<i>E. coli</i>	<i>S. typhi</i>	<i>P. aeruginosa</i>	<i>S. dysenteriae</i>
IL01	10.0 \pm 1.0	5.0 \pm 1.0	7.5 \pm 1.0	4.0 \pm 1.0	4.0 \pm 1.0	6.5 \pm 1.0
IL02	8.0 \pm 1.0	3.0 \pm 1.0	5.5 \pm 1.0	15.0 \pm 1.0	7.5 \pm 1.0	8.0 \pm 1.0
IL03	20.0 \pm 1.0	10.0 \pm 1.0	16.0 \pm 1.0	11.0 \pm 1.0	7.0 \pm 1.0	9.0 \pm 1.0

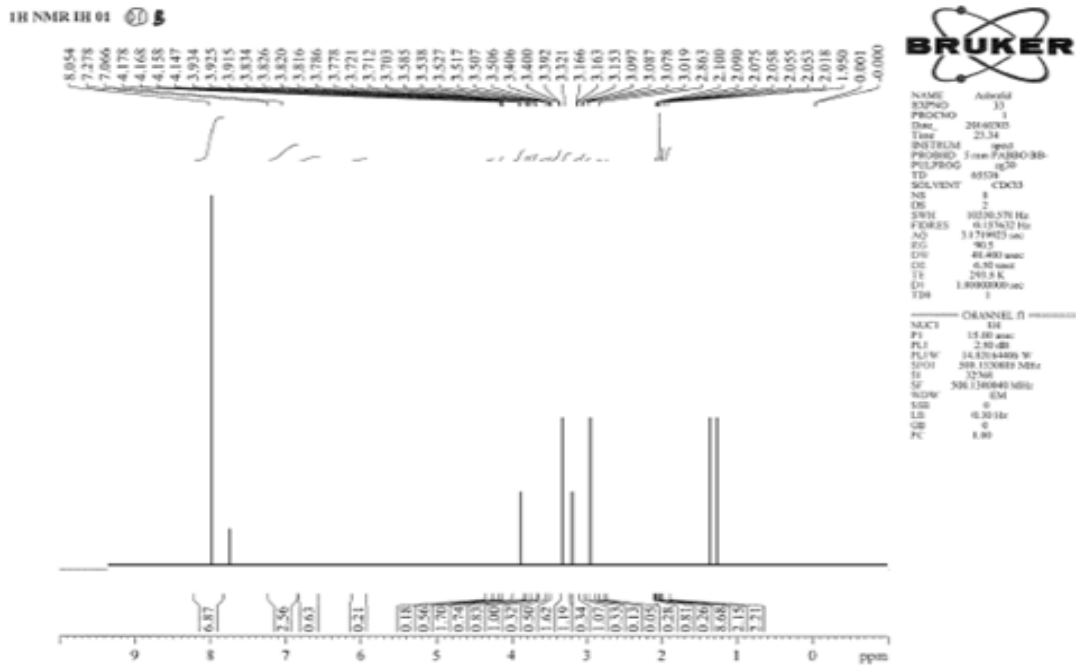


Fig. 7. 1H NMR for II03

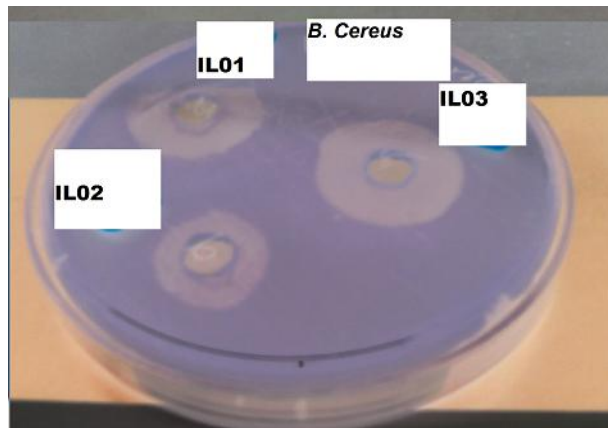


Fig. 8. Zone of inhibition for 20% solution against *B. cereus*

4.3 Determination of MIC

All the data of Table 2 was placed in a graph in excel file. From the data and graph, the MIC can easily determine.

From the graph, it is easy to determine the MIC point for the IL01 sample at which the zone of inhibition is zero. The obtained MIC of *Bacillus cereus*, *Staphylococcus aureus*, *Escherichia coli*, *Salmonella typhi*, *Pseudomonas aeruginosa* and *Shigella dysenteriae* is 250 Mm/L, 250 Mm/L,

75 Mm/L, 250 Mm/L, 250 Mm/L, 75 Mm/L respectively.

From the graph, it is easy to determine the MIC point for the IL02 sample at which the zone of inhibition is zero. The obtained MIC of *Bacillus cereus*, *Staphylococcus aureus*, *Escherichia coli*, *Salmonella typhi*, *Pseudomonas aeruginosa* and *Shigella dysenteriae* is 75.0 Mm/L, 250 Mm/L, 125 Mm/L, 125 Mm/L, 500 Mm/L and 75 Mm/L.

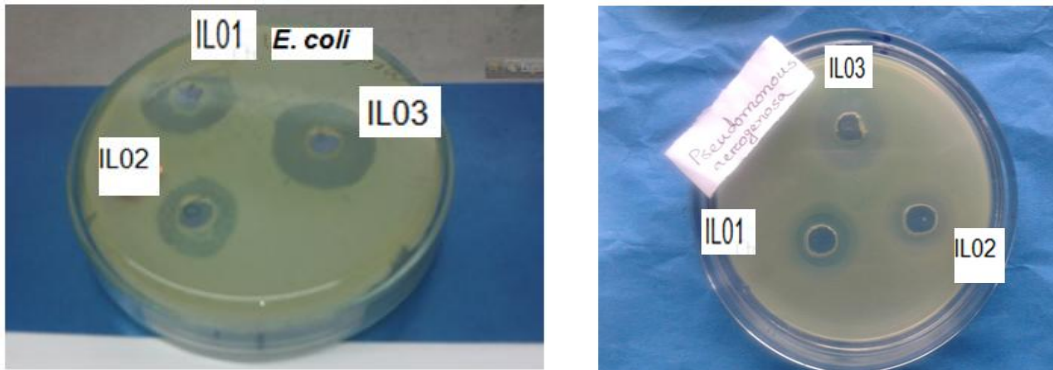


Fig. 9. Zone of inhibition for 20% solution against *E. coli* and *P. aeruginosa*

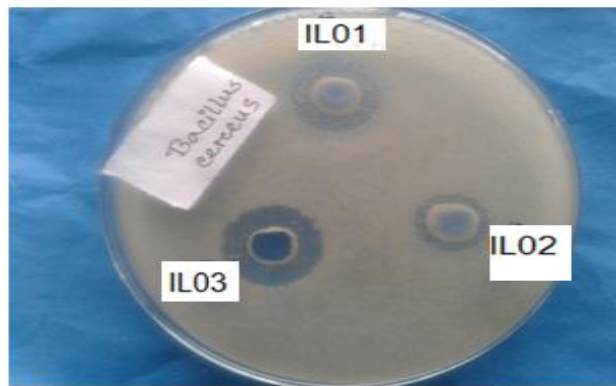


Fig. 10. Zone of inhibition for 500 mM/L solution against *B. cereus*

Table 2. Data of 1000 mM/L, 750 mM/L, 500 mM/L, 250 mM/L, 125 mM/L and 75 mM/L against different Bacteria

		<i>B. cereus</i>	<i>S. aureus</i>	<i>E. coli</i>	<i>S. typhi</i>	<i>P. aeruginosa</i>	<i>S. dysenteriae</i>
L01	1000 mM/L	9.0±1.0	6.0±1.0	11.0±1.0	8.0±1.0	7.0±1.0	10.0±1.0
	750 mM/L	8.0±1.0	4.5±1.0	9.0±1.0	6.0±1.0	5.0±1.0	8.0±1.0
	500mM/L	6.0±1.0	1.5±1.0	6.5±1.0	3.0±1.0	3.0±1.0	5.0±1.0
	250mM/L	0.0	0.0	3.5±1.0	0.0	0.0	3.0±1.0
	125mM/L	0.0	0.0	1.5±1.0	0.0	0.0	1.5±1.0
	75 Mm/L	0.0	0.0	0.0	0.0	0.0	0.0
IL02	1000 mM/L	11.0±1.0	7.0±1.0	8.0±1.0	8.5±1.0	6.0±1.0	12.0±1.0
	750 mM/L	9.5±1.0	5.0±1.0	6.0±1.0	7. ±1.0	2.5±1.0	10.0±1.0
	500mM/L	3.5±1.0	2.0±1.0	3.0±1.0	5.0±1.0	0.0	7.0±1.0
	250mM/L	5.0±1.0	0.0	1.0±1.0	2.0±1.0	0.0	3.0±1.0
	125mM/L	3.0±1.0	0.0	0.0	0.0	0.0	1.5±1.0
	75 Mm/L	0.0	0.0	0.0	0.0	0.0	0.0
IL03	1000 mM/L	17.0±1.0	15. ±1.0	16.0±1.0	17.0±1.0	8.0±1.0	14.0±1.0
	750 mM/L	11.517.0±1.0	12.5±1.0	11.5±1.0	12.0±1.0	3.0±1.0	12.5±1.0
	500mM/L	7.0±1.0	11.0±1.0	7.0±1.0	4.0±1.0	2.5±1.0	11.0±1.0
	250mM/L	4.5 ±1.0	5.5±1.0	3.0±1.0	0.00	0.00	5.0±1.0
	125mM/L	3.0±1.0	0.0	0.00	0.00	0.00	0.00
	75 Mm/L	0.0	0.0	0.00	0.00	0.00	0.00

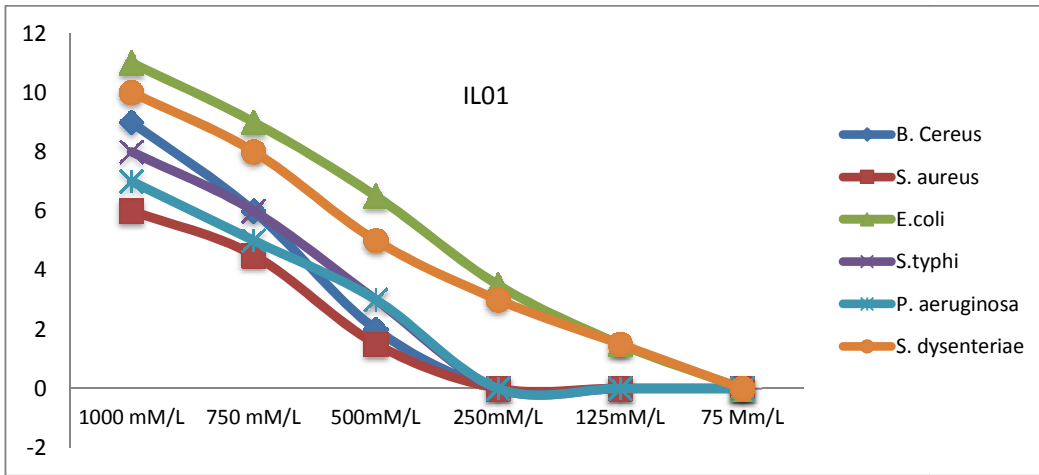


Fig. 11. Determination of MIC for IL01

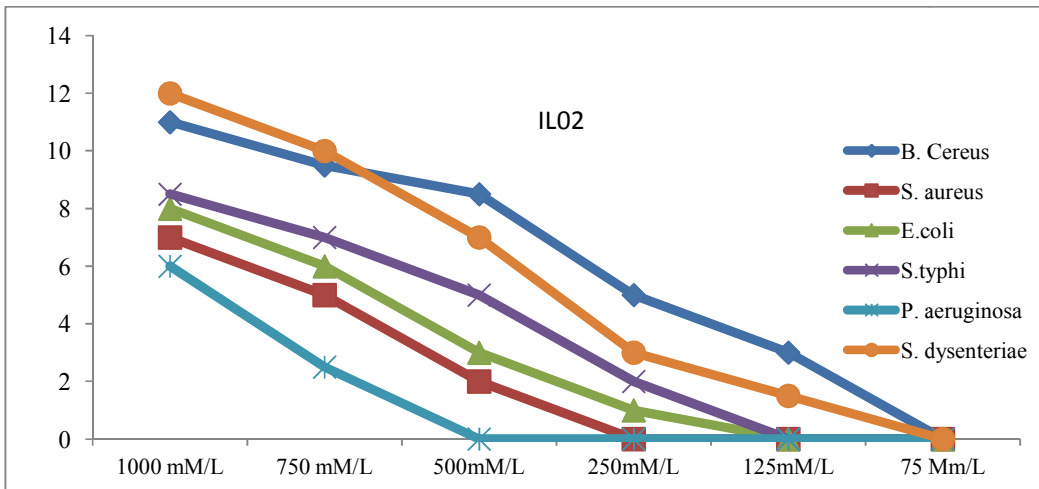


Fig. 4. Determination of MIC for IL02

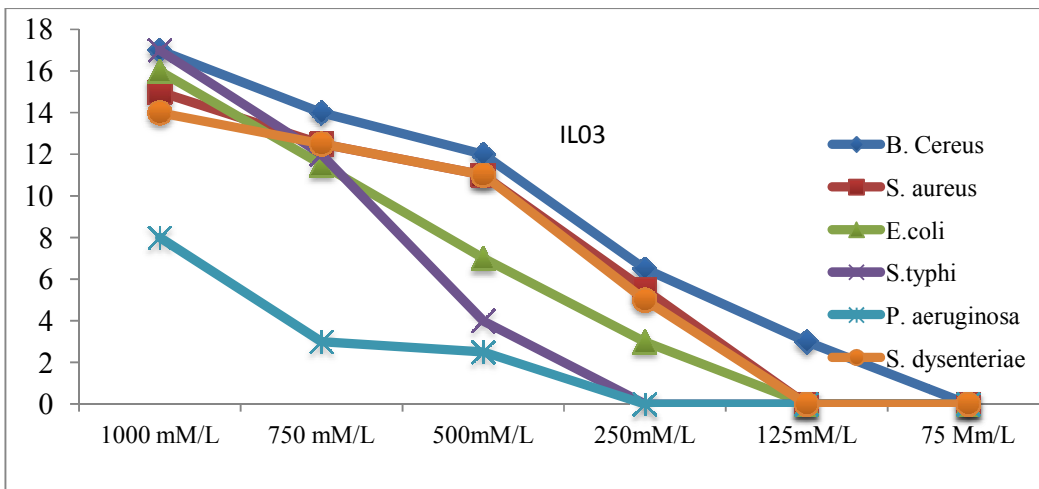


Fig. 5. Determination of MIC for IL03

From the graph, it is easy to determine the MIC point for the IL02 sample at which the zone of inhabitation is zero. The obtained MIC of *Bacillus cereus*, *Staphylococcus aureus*, *Escherichia coli*, *Salmonella typhi*, *Pseudomonas aeruginosa* and *Shigella dysenteriae* is 75 Mm/L, 125 Mm/L, 125 Mm/L, 250 Mm/L, 250 Mm/L and 125 Mm/L.

4.4 Antifungal Screening Test

Aspergillus niger and *Rhizopus azzahra* were used for evaluating the antifungal activity of all synthesized compounds. The antifungal activity was evaluated by Well diffusion method [26]. The media was altered Potato dextrose broth (abbreviated "PDB") is formulated identically to PDA, omitting the agar. Common organisms that can be cultured on PDB are molds such as *Aspergillus niger* and *Rhizopus azzahra*. All synthesized compounds were dissolved in water or methanol basis on their solubility for making the concentration 1000 mM/L. The 100 μ L solution of ILs were taken in petri-plate. The Media of PDB was dispersed and solidified. A well of 5 mm was made in the middle of Petri-plate using cork-borer and the fungal lead was placed there. The plates were then kept in the incubator for 96 h at 37°C. After 3 days, the

fungal growth in presence of ILs was measured and analyzed.

The antifungal test was completed and calculated the growth percentage compared with the control where the growth of control is 100% percent. The growth percentage is deduced by the following equation:

$$\% \text{Growth} = \frac{\text{Growth of fungi with ILs solution}}{\text{Growth of fungi without ILs solution as control}} \times 100$$

4.5 Comparative Study of Toxicity

To compare the antibacterial toxicity, all data was taken in Microsoft Excel to make bar graph.

From the Figs. 7 and 8, it is shown that in all concentration of 1000, 750 and 500 Mm/L, the IL03 can show high antimicrobial activity the IL02 and IL01 respectively. The antimicrobial activity of IL02 is also partially greater than IL01. From this study, it is written that the length of cation has an important rule on bioactivity. With increasing the length and number of alkyl group in cation, the antibacterial activity increases. In the other hand, the antifungal activity is greater in IL01 than IL02 and IL03.

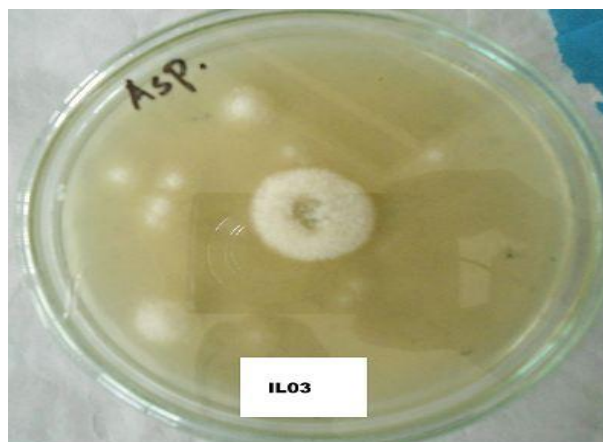


Fig. 6. Zone of inhabitation for antifungal activity of *Aspergillus niger*

Chemicals tested	Zone of growth (in mm)		Percent growth		Percentage of inhabitation	
	<i>Aspergillus niger</i>	<i>Rhizoplus azzahra</i>	<i>Aspergillus niger</i>	<i>Rhizoplus azzahra</i>	<i>Aspergillus niger</i>	<i>Rhizoplus azzahra</i>
Control	28.0 \pm 1.0	31.0 \pm 1.0	100%	100%	0.0%	0.0 %
IL01	17.0 \pm 1.0	21.5 \pm 1.0	60.71%	52.43%	39.29%	47.57%
IL02	26.5 \pm 1.0	26.0 \pm 1.0	92.85%	83.87%	7.15%	16.12%
IL03	24.0 \pm 1.0	25.0 \pm 1.0	85.7%	80.65%	14.30%	19.35%

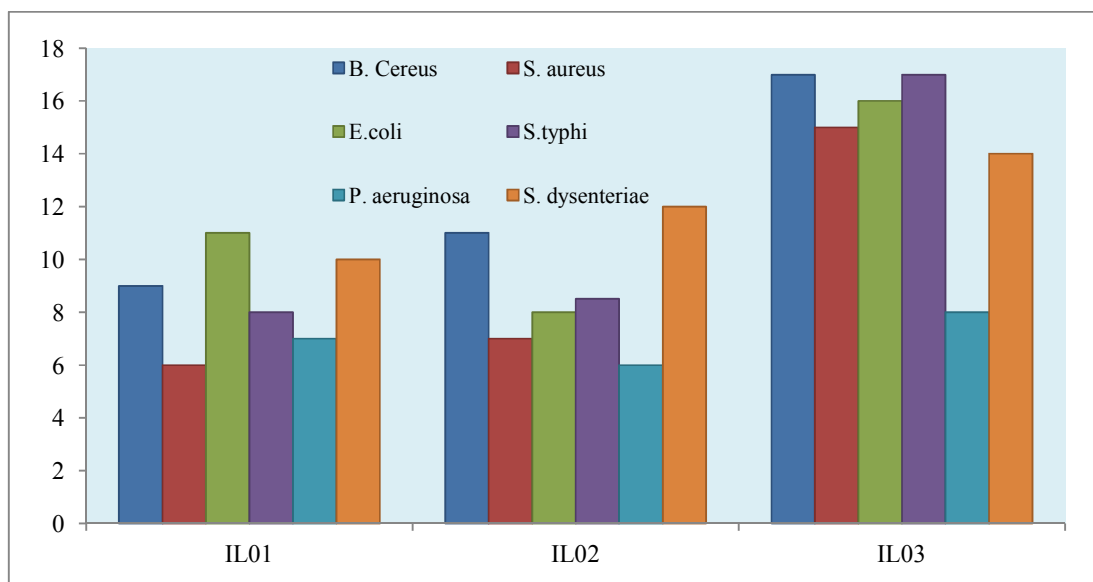


Fig. 7. Comparative antimicrobial studies for 1000 mM/L sample

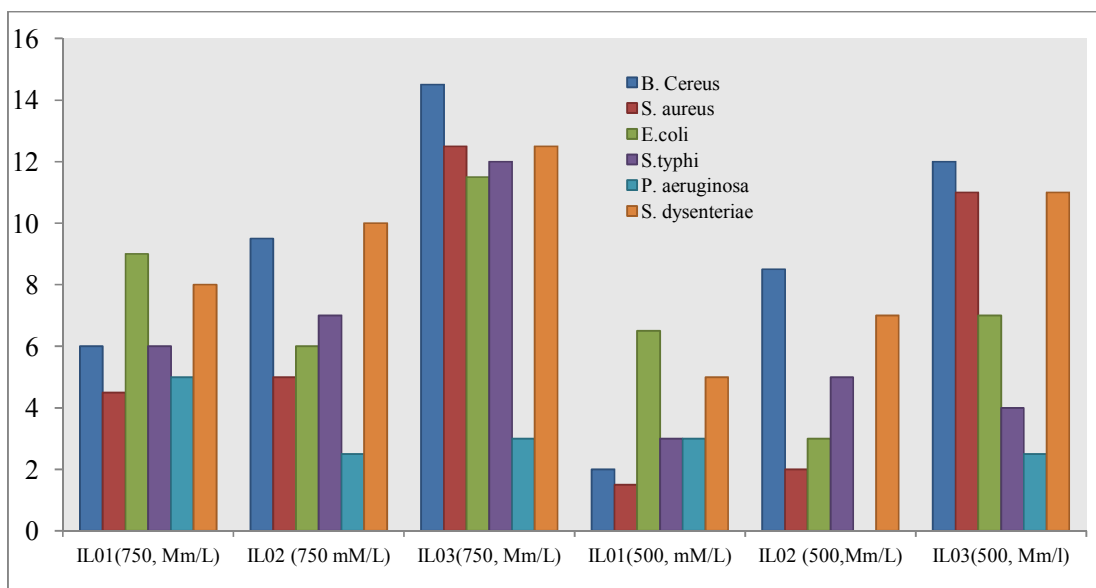


Fig. 8. Comparative antimicrobial studies for 750 mM/L and 500 mM/L sample

5. CONCLUSION

This study showed the three quaternary ammonium ILs and their synthesis with antimicrobial toxicity profiles. The synthesized ILs was characterized by Fourier Transform Infrared Spectroscopy (FT-IR), Nuclear Magnetic Resonance (¹H-NMR) Spectroscopy and elemental analysis. An antibacterial screening was conducted using the well-diffusion technique with ILs solutions of 20% concentrations for initial

screening and show the good response. It is a good effective result for using the bioactive molecule. To establish the bioactivity, the MIC was determined using serial dilution technique and show 250 to 75 Mm/L concentrations. It is different to different pathogens. The IL01 is more antifungal than IL03 and IL02 respectively. All the data of antibacterial and antifungal activity supports that the IL01, IL02, and IL03 are good bioactive synthesis ionic liquids molecules.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Walden P. Molecular weights and electrical conductivity of several fused salts. *Bull. Russian Acad. Sci.* 1914;8:405-422.
2. Wasserscheid P, Keim W. Ionic liquids—new “solutions” for transition metal catalysis. *Angewandte Chemie International Edition.* 2000;39(21):3772-3789.
3. Hasse B, Lehmann J, Assenbaum D, Wasserscheid P, Leipertz A, Fröba A. Viscosity, interfacial tension, density, and refractive index of ionic liquids [EMIM][MeSO₃], [EMIM][MeOHPO₂], [EMIM][OcSO₄], and [BBIM][NTf₂] in dependence on temperature at atmospheric pressure. *Journal of Chemical and Engineering Data.* 2009;54(9):2576-2583.
4. Wang X, HF, Yang M, Melcher B, Fekete M, Mudring AJ, Wasserscheid P, Meyer K. A new class of double alkyl-substituted, liquid crystalline imidazolium ionic liquids - A unique combination of structural features, viscosity effects, and thermal properties *Chemical Communications.* 2009;47:7405-7407.
5. Fröba A, Rausch M, Krzeminski K, Assenbaum D, Wasserscheid P, Leipertz. Thermal conductivity of ionic liquids: Measurement and prediction *International Journal of Thermophysics.* 2010;31(11-12):2059-2077.
6. Plechkova NV, Seddon KR. Applications of ionic liquids in the chemical industry. *Chemical Society Reviews.* 2008;37(1): 123-150.
7. Welton T. Room-temperature ionic liquids. Solvents for synthesis and catalysis. *Chemical Reviews.* 1999;99(8):2071-2084.
8. Wilkes JS, RRD, Seddon KR. Ionic liquids: Industrial applications to green chemistry. *American Chemical Society.* 2002;818: 214–229.
9. Plechkova NV, SKR. Applications of ionic liquids in the chemical industry. *Chemical Society Reviews.* 2008;37:123–150.
10. Carlo Pretti CC, Daniela Pieraccini, Michela Gregori, Francesca Abramo, Gianfranca Monnia, Luigi Intorrec. Acute toxicity of ionic liquids to the zebrafish (Danio rerio). *Green Chemistry.* 2006;8: 238-240.
11. Docherty KCFKJ. Toxicity and antimicrobial activity of imidazolium and pyridinium ionic liquids. *Green Chemistry.* 2005;7:185–189.
12. You Yu YN. Toxicity and antimicrobial activities of ionic liquids with halogen anion. *Journal of Environmental Protection.* 2011;2:298-303.
13. Bicak N. A new ionic liquid: 2-hydroxy ethylammonium formate. *Journal of Molecular Liquids.* 2004;116:37-44.
14. Rajendran A, RD, a.PM. Green synthesis of biologically active pyrazolopyrimidine Derivatives Using an Ionic liquid 2-Methyl-3-butylimidazolium chloride. *International Journal of ChemTech Research.* 2011; 3(1):293-297.
15. Cremer T, Killian M, Gottfried J, Paape N, Wasserscheid P, Maier F, Steinrück HP. Physical vapor deposition of [EMIM][Tf₂N]: A new approach to the modification of surface properties with ultrathin ionic liquid films. *Chem. Phys.* 2008;9(15):2185-2190.
16. Buseti A, Crawford DE, Earle MJ, Gilea M, Gilmore BF, Gorman SP, Lavery G, Lowry AF, McLaughlin M, Seddon KR. Antimicrobial and antibiofilm activities of 1-alkylquinolinium bromide ionic liquids. *Green Chemistry.* 2010;12:420-425.
17. Hough-Troutman WL, SM, Griffin S, Reichert WM, Mirska I, Jodynis-Leibert J, Adamska T, Nawrot J, Stasiewicz M, Rogers RD, Pernak J. Ionic liquids with dual biological function: Sweet and antimicrobial, hydrophobic quaternary ammonium-based salts. *New Journal of Chemistry.* 2009;33:26–33.
18. Hosseinzadeh FM, M, Galehassadi M. Synthesis and characterization of ionic liquid functionalized polymers for drug delivery of an anti-inflammatory drug. *Des. Monomers Polym.* 2012;15:379–388.
19. Hough-Troutman WL, SM, Rodriguez H, Swatloski RP, Spear SK, Daly DT, Pernak J, Grisel JE, Carliss RD, Soutullo MD, Davis JH, Rogers RD. The third evolution of ionic liquids: Active pharmaceutical ingredients. *New Journal of Chemistry.* 2007;31:1429–1436.
20. Yang Z, PW. Ionic liquids: Green solvents for nonaqueous biocatalysis. *Enzyme Microbiology and Technology.* 2005;19–28.
21. Zhao H, SX, Ma P. Review: Use of ionic liquids as green solvents for extractions.

- Journal of Chemical Technology and Biotechnology. 2005;80:1089–1096.
22. Alvarez VH, DN, Gonzáles-Cabaleiro R, Mattedi S, Martin-Pastor M, Iglesias M, Navaza JM. Brønsted ionic liquids for sustainable process: Synthesis and physical properties. Journal of Chemical & Engineering Data. 2010;55:625-632.
23. Iglesias M, GOR, Cota I, Medina F. Brønsted ionic liquids: Study of physico-chemical properties and catalytic activity in aldol condensations. Chemical Engineering Journal. 2010;162:802-808.
24. Bonev B, Hooper J, Parisot J. Principles of assessing bacterial susceptibility to antibiotics using the agar diffusion method. Journal of Antimicrobial Chemotherapy. 2008;61(6): 1295-1301.
25. Standards NCfCL. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard, 5th edn. NCCLS document M7-A5, Wayne, PA; 2000.
26. Standards NCfCL. Reference method for broth dilution antifungal susceptibility testing of yeasts; approved standard. NCCLS document M27-A, Wayne, PA.; 1997.

© 2017 Hossain and Kumer; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
<http://www.sciencedomain.org/review-history/22953>