



Transmission of Anti-SARS-CoV-2 IgG Antibodies to Newborns by Vaccination of Pregnant Women

Jorge Vasconcelos ^{a*}, Ana Furtado Lima ^a, Carlos Ponte ^a
and Rita Pinto ^a

^a Hospital de Divino Espírito Santo, Ponta Delgada, Portugal.

Authors' contributions

This work was carried out in collaboration among all authors. Authors JV and AFL did Project development was performed. Authors JV and RP The data collection was performed. Author JV The analysis of the data was performed. Authors AFL and CP contributed with paper review. All authors read and approved the final manuscript.

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ABSTRACT

Background and Aims: Vaccination in pregnant women is already performed for viruses and bacteria, which has advantages for the mother and the newborn. Besides a general rejection of COVID-19 vaccines in this population, these are considered safe, effective and may prevent severe disease in pregnant women. Neutralizing antibodies titer measured in vaccinated pregnant is highly predictive of immune protection and can have a protective effect on the newborn. This study aimed to confirm the transfer of the antibodies across the placenta and to test if other factors impact the antibodies titer in the newborn circulation.

Study design and methods: We collected 24 samples of umbilical cord blood from women vaccinated during pregnancy or in the three months before pregnancy and analyzed the data of those pregnancies.

Place and Duration of Study: This cohort study was developed in our hospital between March 2021 and May 2022.

*Corresponding author: E-mail: mv.jorge95@gmail.com;

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Results: This study confirms the passage of anti-Spike antibodies to the newborn. When comparing the IgG anti-S titers with the timing of vaccination, the analysis suggests that vaccination during the second or third trimester of pregnancy is more effective than before or in early pregnancy ($p=0,005$). As for the vaccine used ($p=0,23$) and the mode of delivery ($p=0,48$), there were no statistical differences.

Conclusion: We suggest the passage of antibodies during the pregnancy, which appears to be more effective during the second and third trimester of gestation. Despite lack of evidence for how long and how effectively this passive immunity can protect the neonate from infection, vaccination of pregnant women should be considered.

Keywords: Transmission; anti-SARS-CoV-2; antibodies newborns; vaccination pregnant women.

1. INTRODUCTION

In December 2019, uncommon cases of pneumonia of unknown etiology were reported in Wuhan (China). Within a few months, the coronavirus disease 2019 (COVID-19), caused by acute respiratory syndrome coronavirus 2 (SARS-CoV-2), spread rapidly all over the world. This viral infection can cause a mild to severe illness, depending on how immune system encounters the virus [1].

Early on it, was clear that the development of a vaccine would be essential to control the disease, and in less than a year after it was declared a pandemic disease by WHO (World Health Organization) disease, the first vaccines against SARS-CoV-2 were already being distributed around the world [2].

Vaccination in pregnant women is not a new topic as it is already being done for other viruses, like influenza, or bacteria, like Bordetella pertussis. These strategies can have some advantages for the mother, like protection from a severe form of the disease, as well for the newborn, with passive protection from the antibodies produced by pregnant women and transferred to the fetus throughout the pregnancy [3].

Studies have also reported an increased incidence of preeclampsia-like symptoms in infected mothers, such as hypertension, immune dysfunction, and thrombocytopenia without preeclampsia. These complications and the severity of SARS-CoV-2 infection in pregnancy are likely due to the physiological changes in pregnant women, including increased cardiovascular requirements, decreased lung capacity, and immunological changes that are otherwise generally accepted to approximate a mildly immunocompromised state [4].

For COVID-19, some of the first effective vaccines were produced with a theoretically safe

technology for pregnant women. Both the vaccines based on adenovirus vectors, like OxfordAstraZeneca ChAdOx1-S and the vaccines based on messenger RNA, like Pfizer-BioNTech BNT162b2, are secure and pose no risk of pregnant women or fetus infection and so are thought to be safe in this population [5-8]. The mother experiences substantial physiologic and immunologic changes during normal pregnancy to ensure proper fetal growth. 7 These changes include the overexpression in the placenta and fetal organs of the angiotensin-converting enzyme 2 (ACE2) [9].

However, it should be kept in mind that pregnant women were excluded from initial clinical trials for COVID-19 vaccines, limiting the recommendations for to this specific population. This led to an initial rejection of vaccination by the obstetric population, despite current data suggesting that pregnant women may be at increased risk for severe illness, admission to an intensive care unit, mechanical ventilation, and even death when compared with nonpregnant women [10,11]. They are also more likely to deliver pre-term newborns which have higher probability to be admitted to the neonatal intensive care unit [10]. This is highlighted by the Centers for Disease Control and Prevention (CDC), which included pregnancy as a risk factor for severe COVID-19 illness [12]. Due to the significant protection offered by passive immunization acquired through maternal antibodies to the vulnerable newborn and its potential impact on vaccination campaigns, comparing the efficiency of placental antibody transfer in the first, second, and third trimester of infection with SARS-CoV-2 needs to continue to be explored [13].

Therefore, it is of great interest to study the maternal immune response to SARS-CoV-2 vaccination during pregnancy, as well as the extent to which maternal antibodies cross the placenta, to predict the degree of passive

protection acquired by the newborn, and factors that can modulate the effectiveness of that process.

Since neutralizing antibodies titer is already established to be highly predictive of immune protection [14], this study aims primarily to document and confirm the presence of antibodies anti-SARS-CoV-2 in the umbilical cord blood of newborns of mothers vaccinated during pregnancy.

2. METHODS

This cohort study was developed at our center (Hospital of Divino Espírito Santo, São Miguel Island, Azores), between March 2021 and May 2022. Every woman who was vaccinated at least with one dose during the current pregnancy or in the three months before pregnancy, without known previous infection with SARS-CoV-2, and who were admitted to the delivery room in labor, was given the opportunity to participate in the study.

At admission, samples of posterior oropharyngeal and nasopharyngeal secretions were obtained to exclude asymptomatic SARS-CoV-2 infection and positive results were excluded from this study.

At the time of the delivery, an umbilical cord blood sample was collected, regardless of the mode of delivery and the gestational age. Then, the blood samples were tested to determine the circulating IgG anti-S protein level by Chemiluminescence.

Additionally, we tested whether gestational age at vaccination, mode of delivery or vaccine used, had an impact on the titer of circulating antibodies in the newborn circulation. To do so, an independent t-test was performed and p value < 0.05 was considered statistically significant. SPSS version 29.0.2.0 was used for statistical analysis.

This study was approved by the ethics committee of the hospital and all patients gave their written consent to enter the study.

3. RESULTS

We collected 24 samples between May 2021 and March 2022 (Table 1). The characteristics of the population studied are resumed in Table 2.

Pregnant women were on average 33 years and all pregnancies went to term. The mode of

delivery was a vaginal delivery in 15 cases (62,5%), including 3 vacuum deliveries, and a cesarean delivery in the other 9 cases (37,5%).

Most pregnancies included in this study were uneventful. The exceptions were one pregnant woman who developed pre-eclampsia at term, one who was diagnosed with small intestine GIST, and another one with a positive aneuploidy screening (who refused amniocentesis but had an infant without apparent structural anomalies).

In 2 cases, the ChAdOx1 S recombinant vaccine was administered, and in the other 22 cases was administered the BNT162b2 vaccine.

In 95,5% of cases the result was above the cut-off to be considered positive (33.6 BAU/mL). So, there was only one negative case, in which the ChAdOx1 S recombinant vaccine was administered some days after the last menstrual period, having rejected the second dose, due to a pregnancy positive test.

As shown in Table 3, the first and second vaccine doses were administered before conception in one-third and 15,4% of the cases, respectively. The vaccines were administered during different gestational ages in different pregnant women: only 2 doses were administered in the first trimester (none as a first dose); 21 in the second trimester (11 as a first dose); and 12 in the third trimester (5 as a first dose). The latest dose was given at 35 weeks ⁺⁴ days, but nonetheless, had a delay of 25 days between the second dose and the delivery.

There was no statistical difference of the IgG anti-S titers (p=0,48) when compared the result of samples obtained after a vaginal delivery (Median of 1043 BAU/mL ± 801) and after cesarean delivery (Median of 811 BAU/mL ± 634) (Graphic 1). When comparing the vaccine used, there was no statistical difference (p=0,23) when pregnant women were vaccinated with the ChAdOx1 S recombinant vaccine (Median of 352 BAU/mL ± 489) or with Pfizer BNT162b2 vaccine (Median of 1011 BAU/mL ± 739) (Graphic 2).

Regarding the timing of vaccine doses (Graphic 3), there was a statistical difference (p=0,005) between women who were vaccinated in the second or third trimester (Median of 1211 BAU/mL ± 664) and women vaccinated before or very early in the gestation (Median of 163 BAU/mL ± 523). When comparing the titers of antibodies anti-S in the umbilical cord samples, women vaccinated in the 2nd trimester (Median of 864 BAU/mL ± 486) had lower titers (p=0,005)

when comparing with women vaccinated in the 3rd trimester (Median of 1708 BAU/mL \pm 575).

Table 4 resumes the statistical results described here.

4. DISCUSSION

It is established that COVID-19 disease can cause severe disease in pregnant women and can be associated with poor obstetric outcomes. A meta-analysis from 2020 reports a 27% rate of adverse events, with an unusually high cesarean rate [15]. As already mentioned, neonates born from mothers with COVID-19 can also have severe comorbidities like respiratory distress syndrome, pneumonia, and other, secondary to iatrogenic preterm birth and maternal status [16]. Therefore, it is central to mention that vertical transmission can occur, and although the risk is quite low (estimated to be 8% or even lower) [15,17,18], it should not be overlooked. Another consideration is the possibility of postnatal infection by respiratory transmission [19], emphasizing the importance of their previously acquired protection.

Production of IgG antibodies against SARS-CoV-2 virus occurs following natural infection, a process that also occurs with active immunization. Current data suggests that vaccine-induced immune responses are significantly greater than the response that occurs with natural infection [20,21].

Early in the final weeks of the first trimester of gestation, maternal IgG is transferred across the placenta to the developing fetus to protect them from infection as a neonate. This process is called passive immunization and it peaks in the last trimester, where the majority of IgG is transferred in the final 4 weeks of gestation. Although most IgG antibody crosses the placenta in the third trimester, the process is time-dependent, so immunization should be ideally done at least 6 or more weeks prior to delivery [22].

The COVID-19 vaccines that were early commercialized were based on messenger RNA technology that targeted the S protein (protein Spike). Therefore, the circulating level of IgG anti-S is a measure of the immune response to the vaccination [23]. Although this response does not only rely on the titer of IgG anti-S antibodies (since there is also a cellular immune response), this is an objective measure of the reactivity of the immune system and a way to

predict an eventual passive protection of the newborn. As evidence supports, anti-SARS-CoV-2 antibodies titers are associated with a lower risk of symptomatic disease [21,24,25].

In agreement with other recent studies, it was observed that the anti-SARS-CoV-2 antibodies, produced by the pregnant woman's immune system in response to the vaccine, crossed the placenta and became present in the fetus's blood resulting in passive immunity, at least in the neonatal period [24,26,27].

The increasing levels of maternal IgG and placental IgG transfer ratio over time suggest that the timing between vaccination and delivery may be an important factor to consider in vaccination strategies in this population.

Although some studies suggest that a longer latency between vaccination and delivery correlates with higher anti-S IgG titers [26,28], there is already evidence that vaccination of pregnant women in the third trimester can be equally effective [29]. In fact, these vaccines induce a rapid stimulation of the immune system with antibody production as early as 5 days after the first dose, and transplacental transfer of passive immunity to the neonate as early as 16 days after the first vaccination dose [30,31]. As we demonstrate, vaccination in the third trimester seems to correlate to higher titers of anti-Spike antibodies in the umbilical cord blood at the delivery.

We also tried to prove if there was any correlation between the anti-S antibodies titers and the type of vaccine used or the mode of delivery. The mode of delivery did not seem to interfere with the anti-S titers in the umbilical cord as that passage occurs during gestation and not at the time of the delivery. Considering the type of vaccine used, although the only negative sample was from a pregnant woman vaccinated with the ChAdOx1 S recombinant vaccine, she received only one dose, instead of the usual two. Besides that, our sample was too small to draw a conclusion based on the vaccines used.

This study has some limitations, including that it did not look at the long-term effects of vaccines on newborns and mothers. It would be also interesting to address a proposed preventive chemotherapy for COVID-19, such as ivermectin, and understand who it would impact the benefit of vaccination [32-33].

Table 1. Description of the cases included in the cohort

| | Mother Age (years) | Complications during pregnancy | Gestational age at birth (weeks) | Mode of Delivery | Gestational age at vaccination (weeks) | | Vaccine administered | IgG anti-S spike |
|----|--------------------|--------------------------------|----------------------------------|------------------|--|----------------------|-----------------------|------------------|
| | | | | | 1 st dose | 2 nd dose | | |
| 1 | 36 | None | 40+2 | Vaginal | 20+5 | 23+5 | Pfizer BNT 162b2 | 703 BAU/mL |
| 2 | 27 | None | 38+4 | Vaginal | 28+6 | 31+6 | Pfizer BNT 162b2 | >2080 BAU/mL |
| 3 | 28 | None | 39+0 | Cesarean | PC | 3 | Pfizer BNT 162b2 | 98.6 BAU/mL |
| 4 | 32 | None | 39+1 | Vaginal | 1 | 4 | Pfizer BNT 162b2 | 1500 BAU/mL |
| 5 | 31 | None | 40+6 | Cesarean | 30+3 | 33+3 | Pfizer BNT 162b2 | >2080 BAU/mL |
| 6 | 36 | None | 40+2 | Vaginal | 30+5 | 33+5 | Pfizer BNT 162b2 | >2080 BAU/mL |
| 7 | 38 | None | 39+1 | Vaginal | 32+4 | 35+4 | Pfizer BNT 162b2 | 676 BAU/mL |
| 8 | 33 | None | 39+1 | Vaginal | PC | PC | Pfizer BNT 162b2 | 259 BAU/mL |
| 9 | 31 | None | 39+4 | Vaginal | PC | PC | Pfizer BNT 162b2 | 163BAU/mL |
| 10 | 37 | None | 40+0 | Vaginal | 23+1 | 26+1 | Pfizer BNT 162b2 | 335 BAU/mL |
| 11 | 31 | None | 38+3 | Vaginal | 20+5 | 23+5 | Pfizer BNT 162b2 | 704 BAU/mL |
| 12 | 37 | None | 37+3 | Vaginal | 24+5 | 27+6 | Pfizer BNT 162b2 | >2080 BAU/mL |
| 13 | 43 | Small intestine GIST | 39+0 | Vaginal | 23+6 | 27+6 | Pfizer BNT 162b2 | 430 BAU/mL |
| 14 | 39 | None | 38+6 | Cesarean | PC | PC | Pfizer BNT 162b2 | 59.40 BAU/mL |
| 15 | 37 | None | 39+2 | Cesarean | 23+0 | 26+0 | Pfizer BNT 162b2 | 765 BAU/mL |
| 16 | 31 | None | 38+4 | Cesarean | PC | PC | Pfizer BNT 162b2 | 260 BAU/mL |
| 17 | 31 | None | 38+0 | Cesarean | 23+0 | 26+0 | Pfizer BNT 162b2 | 784 BAU/mL |
| 18 | 29 | None | 39+5 | Vaginal | PC | NA | ChAdOx1 S recombinant | 6.33 BAU/mL |
| 19 | 28 | None | 39+0 | Cesarean | 22+0 | 25+0 | Pfizer BNT 162b2 | 1080 BAU/mL |
| 20 | 28 | Pre-eclampsia at term | 37+1 | Cesarean | 27+4 | 30+0 | Pfizer BNT 162b2 | 1110 BAU/mL |
| 21 | 33 | None | 40+0 | Vaginal | PC | 23+2 | ChAdOx1 S recombinant | 698 BAU/mL |
| 22 | 32 | Positive aneuploidy screening | 39+2 | Cesarean | 22+3 | 25+3 | Pfizer BNT 162b2 | 1060 BAU/mL |
| 23 | 32 | None | 40+3 | Vaginal | 30+5 | 33+6 | Pfizer BNT 162b2 | 1880 BAU/mL |
| 24 | 30 | None | 39+5 | Vaginal | 26+6 | 29+6 | Pfizer BNT 162b2 | 2050 BAU/mL |

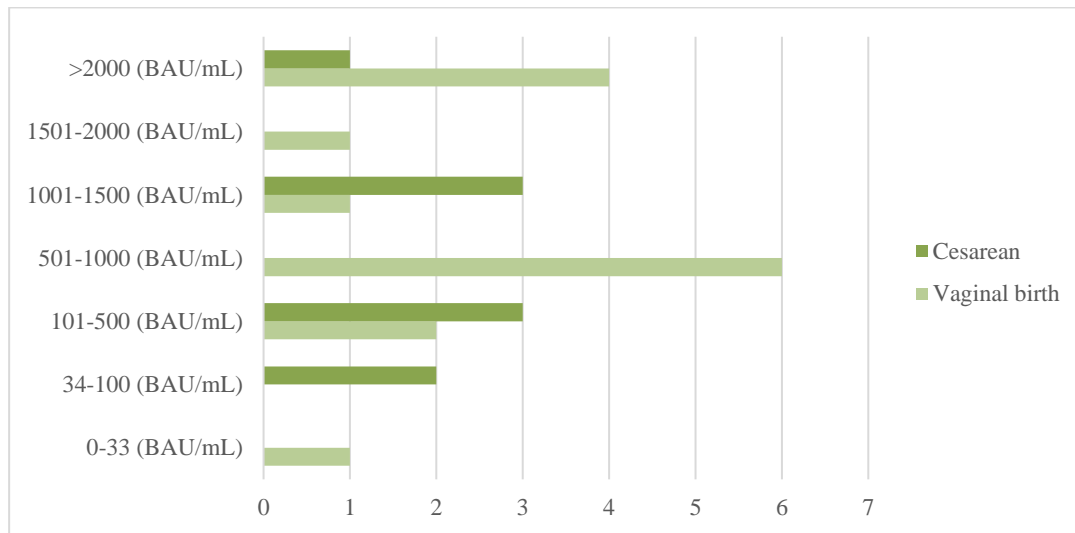
Abbreviations: PC, Preconception; NA, not applicable; GITS, Gastrointestinal Stromal Tumor.

Table 2. Median characteristics of the population studied

| Characteristics of the population studied | Median | |
|---|---------------------|------------|
| Age (years) | 33 (28;43) | |
| Gestational age at birth (week) | 39 + 1 (37+1; 40+6) | |
| Mode of delivery | Vacuum delivery | 3 (12,5%) |
| | Euthocic delivery | 12 (50%) |
| | Cesarian delivery | 9 (37,5%) |
| IgG anti spike | Positive result | 23 (95,8%) |
| | Negative result | 1 (4,2%) |

Table 3. Timing of vaccine doses

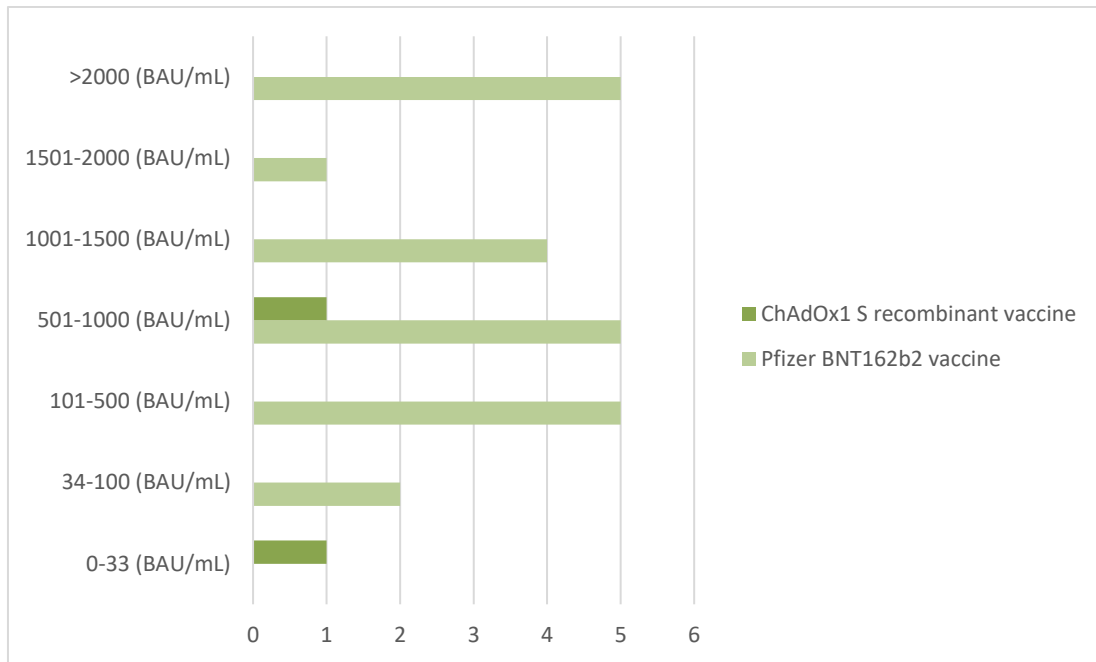
| First Dose (n=24) | |
|---------------------------|------------|
| Before pregnancy | 8 (33,3%) |
| 1º Trimester | 0 (0%) |
| 2º Trimester | 11 (45,8%) |
| 3º Trimester | 5 (20,8%) |
| Second Dose (n=23) | |
| Before pregnancy | 4 (15,4%) |
| 1º Trimester | 2 (8,7%) |
| 2º Trimester | 10 (43,5%) |
| 3º Trimester | 7 (30,4%) |



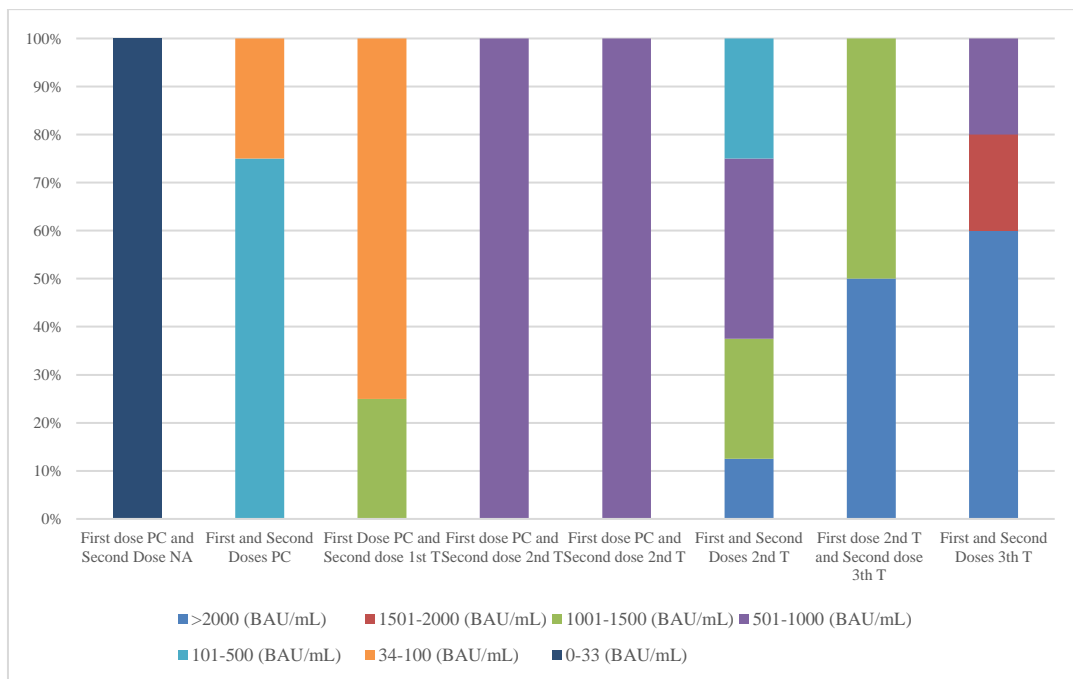
Graphic 1. Anti-Spike titers according to the mode of delivery

Despite the lack of evidence of how long this passive immunity can protect the neonate from infection and how effective is that protection, the confirmation of the passage of these antibodies through the placenta and the fact, already established, that pregnant women are a group of higher risk for severe disease when compared with the general population, may justify vaccination even during pregnancy.

There is still more investigation work to do in this area to help to understand the better dose schedule, if there are differences in the vaccines used, the impact on subsequent pregnancies, the neonatal outcomes and the duration of passive immunity. This would make vaccination more secure and help practitioners to better counsel their patients.



Graphic 2. Anti-Spike titers according to the type of vaccine used



Graphic 3. Anti-Spike titers according to the timing of the vaccine doses
 Abbreviations: PC, Preconception; NA, not applicable; T, Trimester.

Table 4. Effect of mode of delivery, the timing of vaccination, and vaccine used on IgG anti-Spike titers

| Mode of delivery | |
|---------------------------------------|-----------------------------|
| Vaginal delivery | Median of 1043 BAU/mL ± 801 |
| Cesarean delivery | Median of 811 BAU/mL ± 634 |
| p= 0,4671 | |
| Timing of vaccination | |
| Second or third trimester | Median of 1211 BAU/mL ± 664 |
| Before or very early on the gestation | Median of 163 BAU/mL ± 523 |
| p= 0,0051 | |
| Third Trimester | Median of 1708 BAU/mL ± 575 |
| Before pregnancy | Median of 149 BAU/mL ± 115 |
| p= 0,0005 | |
| Third Trimester | Median of 1708 BAU/mL ± 575 |
| Second trimester | Median of 864 BAU/mL ± 486 |
| p= 0,0052 | |
| During pregnancy | Median of 1168 BAU/mL ± 680 |
| Before the pregnancy | Median of 149 BAU/mL ± 115 |
| p= 0,0034 | |
| Vaccine used | |
| Pfizer BNT 162b2 vaccine | Median of 1011 BAU/mL ± 739 |
| ChAdOx1 S recombinant vaccine | Median of 352 BAU/mL ± 489 |
| p= 0,2342 | |

5. CONCLUSION

In conclusion, the use of anti-COVID-19 vaccines should be encouraged and any misconceptions about risks for the mother and the fetus should be clarified considering the evidence available. This work suggests that vaccination during the second or third trimester of gestation is effective than vaccination earlier in pregnancy.

DISCLAIMER (ARTIFICIAL INTELLIGENCE)

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of manuscripts.

CONSENT

All participants included in the study sign an informed consent stating their consent to participate in this study.

ETHICAL APPROVAL

This study was approved by the ethics committee of our institution (with the approval code: Ref.^a 1166/CES/2021). The procedures used in this study are in line with the principles of the Declaration of Helsinki.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Wang C, Horby PW, Hayden FG, Gao GF. A novel coronavirus outbreak of global health concern. *Lancet.* 2020; 395(10223):470–473. . Available:[https://doi.org/10.1016/S0140-6736\(20\)30185-9](https://doi.org/10.1016/S0140-6736(20)30185-9)
2. Chakraborty C, Bhattacharya M, Dhama K. SARS-CoV-2 vaccines, Vaccine Development Technologies, and Significant Efforts in Vaccine Development during the Pandemic: The Lessons Learned Might Help to Fight against the Next Pandemic. *Vaccines.* 2023;11:682. Available:<https://doi.org/10.3390/vaccines11030682>
3. Nassar AH, Hobeika E, Chamsy D, El-Kak F, Usta IM. Vaccination in pregnancy. *Int J Gynaecol Obstet.* 2023; 162(1):18-23. Available:<https://doi.org/10.1002/ijgo.14876>
4. Marchand G, Masoud AT, Grover S, King A, Brazil G, Ulibarri H, Parise J, Arroyo A, Coriell C, Goetz S, Moir C. Maternal and neonatal outcomes of COVID-19

- vaccination during pregnancy, a systematic review and meta-analysis. *npj Vaccines*. 2023;8(1):103.
5. Lopez Bernal J, Andrews N, Gower C, Robertson C, Stowe J, Tessier E et al. Effectiveness of the Pfizer-BioNTech and Oxford-AstraZeneca vaccines on covid-19 related symptoms, hospital admissions, and mortality in older adults in England: test negative case-control study. *BMJ*. 2021;373:n1088. Available:<https://doi.org/10.1136/bmj.n1088>
 6. Leik NKO, Ahmedy F, Guad RM, & Baharuddin DMP. Covid-19 vaccine and its consequences in pregnancy: Brief review. *Annals of medicine and surgery*. 2021;72:103103. Available:<https://doi.org/10.1016/j.amsu.2021.103103>
 7. Bookstein Peretz S, Regev N, Novick L, Nachshol M, Goffer E, Ben-David A et al. Short-term outcome of pregnant women vaccinated with BNT162b2 mRNA COVID-19 vaccine. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology* 2021; 58(3): 450–456. Available:<https://doi.org/10.1002/uog.23729>
 8. Ciapponi A, Bardach A, Mazzoni A, Alconada T, Anderson SA, Argento FJ et al. Safety of components and platforms of COVID-19 vaccines considered for use in pregnancy: A rapid review. *Vaccine*. 2021; 39(40):5891–5908. Available:<https://doi.org/10.1016/j.vaccine.2021.08.034>
 9. Martin-Vicente M, Carrasco I, Muñoz-Gomez MJ, Lobo AH, Mas V, Vigil-Vázquez S, Vázquez M, Manzanares A, Cano O, Alonso R, Sepúlveda-Crespo D. Antibody levels to SARS-CoV-2 spike protein in mothers and children from delivery to six months later. *Birth*. 2023; 50(2):418-27.
 10. Ilotey J, Stallings E, Bonet M, Yap M, Chatterjee S, Kew T et al. Clinical manifestations, risk factors, and maternal and perinatal outcomes of coronavirus disease 2019 in pregnancy: living systematic review and meta-analysis. *BMJ*. 2020; 370:m3320. Available:<https://doi.org/10.1136/bmj.m3320>
 11. Celewicz A, Celewicz M, Michalczyk M, Wozniakowska-Gondek P, Krejczyk K, Misiek M, Rzepka R. Pregnancy as a risk factor of severe COVID-19. *J clin Med*. 2021; 10, 5458. Available:<https://doi.org/10.3390/jcm10225458>
 12. CDC Underlying Medical Conditions Associated with Higher Risk for Severe COVID-19: Information for Healthcare Professionals. Updated Feb 9, 2023. Available:<https://www.cdc.gov/coronavirus/2019-ncov/hcp/clinicalcare/underlyingconditions.html>. [Accessed on 29, April, 2023]
 13. Lucot-Royer L, Nallet C, Vouga M, Puyraveau M, Mauny F, Marty-Quinternet S, Bertholdt C, Bory JP, Devalland C, Canaguier M, Copolla C. Analysis of the transplacental transmission of SARS CoV-2 virus and antibody transfer according to the gestational age at maternal infection. *Scientific Reports*. 2024;14(1):3458.
 14. Khoury DS, Cromer D, Reynaldi A, Schlub TE, Wheatley AK, Juno JA et al. Neutralizing antibody levels are highly predictive of immune protection from symptomatic SARS-CoV-2 infection. *Nature medicine*. 2021; 27(7):1205–1211. Available:<https://doi.org/10.1038/s41591-021-01377-8>
 15. Dubey P, Reddy SY, Manuel S, & Dwivedi AK. Maternal and neonatal characteristics and outcomes among COVID-19 infected women: An updated systematic review and meta-analysis. *European journal of obstetrics, gynecology, and reproductive biology*. 2020; 252:490–501. Available:<https://doi.org/10.1016/j.ejogrb.2020.07.034>
 16. Kyle MH, Glassman ME, Khan A, Fernández CR, Hanft E, Emeruwa UN et al. A review of newborn outcomes during the COVID-19 pandemic. *Seminars in perinatology*. 2020; 44(7):151286. Available:<https://doi.org/10.1016/j.semperi.2020.151286>
 17. Gajbhiye RK, Modi DN, Mahale SD. Pregnancy outcomes, Newborn complications and Maternal-Fetal Transmission of SARS-CoV-2 in women with COVID-19: A systematic review of 441 cases. *medRxiv*. 2020;7(2):21-28. Available:<https://doi.org/10.1101/2020.04.11.20062356>
 18. Fenizia C, Biasin M, Cetin I, Vergani P, Mileto D, Spinillo A et al. Analysis of SARS-CoV-2 vertical transmission during

- pregnancy. *Nature communications*. 2020;11(1):5128.
Available:<https://doi.org/10.1038/s41467-020-18933-4>
19. Novoa RH, Quintana W, Llancaí P, Urbina-Quispe K, Guevara-Ríos E, & Ventura W. Maternal clinical characteristics and perinatal outcomes among pregnant women with coronavirus disease 2019. A systematic review. *Travel medicine and infectious disease*. 2021;39:101919. Available:<https://doi.org/10.1016/j.tmaid.2020.101919>
 20. Flannery DD, Gouma S, Dhudasia MB, Mukhopadhyay S, Pfeifer MR, Woodford EC et al. Assessment of Maternal and Neonatal Cord Blood SARS-CoV-2 Antibodies and Placental Transfer Ratios. *JAMA pediatrics*. 2021;175(6):594–600. Available:<https://doi.org/10.1001/jamapediatrics.2021.0038>
 21. Gray KJ, Bordt EA, Atyeo C, Deriso E, Akinwunmi B, Young N et al. COVID-19 vaccine response in pregnant and lactating women: A cohort study. *medRxiv : the preprint server for health sciences*. 2021;03.07.21253094. Available:<https://doi.org/10.1101/2021.03.07.21253094>
 22. Munoz FM, Jamieson DJ. Maternal Immunization. *Obstetrics and gynecology* . 2019; 133(4):739–753. Available:<https://doi.org/10.1097/AOG.0000000000003161>
 23. Walls AC, Park YJ, Tortorici MA, Wall A, McGuire AT, & Velesler D. Structure, Function, and Antigenicity of the SARS-CoV-2 Spike Glycoprotein. *Cell*. 2020;181(2):281–292.e6. Available:<https://doi.org/10.1016/j.cell.2020.02.058>
 24. Cicalini I, Rossi C, Natale L, Cufaro MC, Catitti G, Vespa S et al. Passive Immunity to SARS-CoV-2 at Birth Induced by Vaccination in the First Trimester of Pregnancy. *International journal of environmental research and public health*. 2021;18(23):12789. Available:<https://doi.org/10.3390/ijerph182312789>
 25. Feng S, Phillips DJ, White T, Sayal H, Aley PK, Bibi S et al. Correlates of protection against symptomatic and asymptomatic SARS-CoV-2 infection. *Nature medicine*. 2021;27(11):2032–2040. Available:<https://doi.org/10.1038/s41591-021-01540-1>
 26. Trostle ME, Agüero-Rosenfeld ME, Roman AS, & Lighter JL. High antibody levels in cord blood from pregnant women vaccinated against COVID-19. *American journal of obstetrics & gynecology MFM*. 2021; 3(6):100481. Available:<https://doi.org/10.1016/j.ajogmf.2021.100481>
 27. Zdanowski W, Waśniewski T. Evaluation of SARS-CoV-2 Spike Protein Antibody Titers in Cord Blood after COVID-19 Vaccination during Pregnancy in Polish Healthcare Workers: Preliminary Results. *Vaccines*. 2021;9(6):675. Available:<https://doi.org/10.3390/vaccines9060675>
 28. Mithal LB, Otero S, Shanes ED, Goldstein JA, & Miller ES. Cord blood antibodies following maternal coronavirus disease 2019 vaccination during pregnancy. *American journal of obstetrics and gynecology*. 2021;225(2):192–194. Available:<https://doi.org/10.1016/j.ajog.2021.03.035>
 29. Beharier O, Plitman Mayo R, Raz T, Nahum Sacks K, Schreiber L, Suissa-Cohen Y et al. Efficient maternal to neonatal transfer of antibodies against SARS-CoV-2 and BNT162b2 mRNA COVID-19 vaccine. *The Journal of clinical investigation*. 2021; 131(13): e150319. Available:<https://doi.org/10.1172/JCI150319>
 30. Prabhu M, Murphy EA, Sukhu AC, Yee J, Singh S, Eng D. Antibody Response to Coronavirus Disease 2019 (COVID-19) Messenger RNA Vaccination in Pregnant Women and Transplacental Passage Into Cord Blood. *Obstetrics and gynecology*. 2021;138(2):278–280. Available:<https://doi.org/10.1097/AOG.0000000000004438>
 31. Rottenstreich A, Zarbiv G, Oiknine-Djian E, Zigron R, Wolf DG, & Porat S. Efficient Maternofetal Transplacental Transfer of Anti- Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Spike Antibodies After Antenatal SARS-CoV-2 BNT162b2 Messenger RNA Vaccination. *Clinical infectious diseases: an official publication of the Infectious Diseases Society of America*. 2021;73(10):1909–1912. Available:<https://doi.org/10.1093/cid/ciab266>
 32. Hegazy AA, Alghamdi MS, Shouman WM, Hegazy RA. Mass chemoprophylaxis with

- ivermectin against COVID-19 pandemic: review and authors' perspective. Acta Scientific Medical Sciences. 2021;5(6):47-51.
Available:<https://doi.org/10.31080/ASMS.2020.05.0921>
33. Shoumann WM, Hegazy AA, Nafae RM, Ragab MI, Samra SR, Ibrahim DA, Al-Mahrouky TH, Sileem AE. Use of Ivermectin as a Potential Chemoprophylaxis for COVID-19 in Egypt: A Randomized Clinical Trial. Journal of Clinical & Diagnostic Research. 2021;15(2):27-32.
Available:<https://doi.org/10.7860/JCDR/2021/46795.14529>

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