

Journal of Pharmaceutical Research International

33(62A): 387-395, 2021; Article no.JPRI.77808 ISSN: 2456-9119 (Past name: British Journal of Pharmaceutical Research, Past ISSN: 2231-2919, NLM ID: 101631759)

Evaluation of Antitumour Activity of Ethanolic Extract from *Tribulus terrestris* in Human Breast Cancer Cells

M. Kamalli ^{a≡}, S. Raghunandhakumar ^{b*}^o, D. Ezhilarasan ^{ao} and T. Lakshmi ^{c#}

 ^a Department of Pharmacology, Saveetha Dental College, Saveetha Institute of Medical and Technical Sciences, No.162, PH Road, Chennai, Tamil Nadu, 600 077, India.
 ^b Cancer and Stem Cell Research Laboratory, Department of Pharmacology, Saveetha Dental College, Saveetha Institute of Medical and Technical Sciences, No.162, PH Road, Chennai, Tamil Nadu, 600 077, India.

^c Department of Pharmacology, Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technical Sciences (SIMATS), Saveetha University, Chennai - 600077, India.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JPRI/2021/v33i62A35613

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/77808

Original Research Article

Received 22 October 2021 Accepted 27 December 2021 Published 28 December 2021

ABSTRACT

Introduction: Breast cancer is the form of cancer that occurs in the breast cells. After skin cancer, breast cancer is considered to be the most common cancer diagnosed in women. The number of deaths associated with this disease was increased in case of lack of early detection. *Tribulus terrestris* is an annual herb belonging to the Zygophyllaceae family. Flavonoids, alkaloids, and saponins are some of the main phytoconstituents of the herb. They are known for their pharmacological actions such as anti carcinogenic, anti inflammatory, antimicrobial, antioxidant properties.

Aim: The aim of the present study is to evaluate the antitumor activity of ethanolic extract from *Tribulus terrestris* in MCF-7 human breast cancer cells.

Materials and Methods: The effect of Tribulus terrestris on cell viability was measured by MTT

[■] Graduate;

^o Associate Professor;

[#] Professor;

^{*}Corresponding author: E-mail: raghunandhakumars.sdc@saveetha.com;

assay carried against breast cancer cells and morphological changes were investigated with phase contrast microscopy to confirm its antitumor activity.

Results: the cell viability assay results indicate that 24hrs treatment with ethanolic extract of significantly reduces the cell viability in dose dependent manner. At 40 μ g/ml of the *Tribulus terrestris* extract inhibits 50% cell viability and it has been fixed as IC₅₀ value for further *Tribulus terrestris* experiments.

Conclusion: From the results, the extracts were cytotoxic to the human breast cancer cell and it might be a good therapeutic value for further investigations needed to understand the mechanisms to develop antitumor agents.

Keywords: Anti-tumour; MTT assay; Tribulus terrestris; Breast cancer; Cytotoxicity.

1. INTRODUCTION

Breast cancer is a kind of cancer that develops in the breast cells. Breast cancer is the second most prevalent cancer diagnosed in women, after skin cancer. In the case of early discovery, the number of deaths connected with this condition is constantly decreasing. In the last decade, there has been significant progress in both the knowledge of breast cancer and the development of preventative therapies [1]. Amongst all the malignant cancer types, breast cancer is considered as one of the main causes of death in post menopausal women for 23% of all cancer deaths [2]. Screening mammography helps to identify breast cancer at prior stages of the disease [3]. Breast cancer develops in the lobules or ducts. It can also arise in the breast's fibrous connective tissue or fatty tissue. Lymph nodes serve as a main route for cancer to spread to other regions of the body. Breast cancer symptoms include lumps, thickening, swelling, discomfort, dimpling, redness, and flaky skin. Breast cancer is classified into four types: ductal carcinoma in situ, invasive ductal carcinoma, inflammatory breast cancer, and metastatic breast cancer. Breast cancer is caused mostly by reproductive and hormonal causes. Other variables that contribute to breast cancer include ionising radiation exposure and a hereditary susceptibility. Long-term exposure to high levels of endogenous estrogens raises the risk of menopausal breast cancer in and postmenopausal women [4].

Tribulus terrestris is an annual herb belonging to the Zygophyllaceae family. Flavonoids, alkaloids, and saponins are some of the main phytoconstituents of the herb. They are known for their pharmacological actions such as anticarcinogenic, anti-inflammatory, antimicrobial, antioxidant properties. The fruits and roots of the plant is constantly used as a folk medicine for more than thousand years [5]. The preparations from the plant parts are specifically popular for health issues and diseases such as hormonal imbalance, sexual problems and other heart diseases [6]. In recent years, the beneficial effects of the plant have been found and the use of the plant is gradually increasing in developing countries [7].

Ethanolic extract is considered as a single stream process which can be done under both warm and cold conditions. It can be used as a solvent of extraction at room temperature or under supercooled temperature. Different levels of ethanolic extract of *Tribulus terrestris* were tested to enhance reproductive performance and to reduce hormonal imbalance [8]. Our team has extensive knowledge and research experience that has translate into high quality publications [9-42]. The aim of the study is to evaluate the antitumor activity of ethanolic extract from *Tribulus terrestris* using MCF-7 human breast cancer cells.

2. MATERIALS AND METHODS

2.1 Chemicals

Dulbecco's Modified Eagle Medium (DMEM) medium, 0.25% Trypsin-EDTA solution, sodium bicarbonate solution, bovine serum albumin (BSA), low melting agarose, MTT (3-(4,5dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) from Sigma Chemicals Co., St. Louis, USA. Fetal bovine serum (FBS) and antibiotic/antimycotic solution. Dimethyl sulfoxide (DMSO) were from Himedia, Sodium phosphate monobasic and dibasic, sodium chloride, sodium hydroxide, sodium carbonate, hydrochloric acid and methanol were purchased from Sisco Research Laboratories (SRL) India.

2.2 Preparation of Ethanolic Plant Extract

The whole plant of *Tribulus terrestris* powder was commercially purchased from The Indian Medical

Practitioners Co-operative Pharmacy and Stores Ltd (IMPCOPS), (Chennai, India) for the present study. 100gm of powder with 500ml of 95% ethanol mixed well and kept at room temperature for 3 days in a static condition. Then the extract was filtered and fixed into a soxhlet apparatus subjected to evaporation at room temperature till a semisolid mass and further it was concentrated in vacuum evaporate and immediately stored at 4°C for further experiment.

2.3 Cell Line and Culture

The Human breast cancer cells, MCF-7 was procured from National Centre for Cell Science (NCCS) Pune, India. The cells were grown in T25 culture flasks containing DMEM high glucose medium provided with 10% FBS, Penicillin (100 IU/ml), Streptomycin (100 µg/ml) and Amphotericin B along with 7.5% sodium bicarbonate and incubated at 37°C in 5% CO₂ incubator. After 3 days, about 80-90% confluent monolayer (adherent) formation was confirmed by inverted microscope and it was carried for further experiment by using Trypsin-EDTA solution.

2.4 Cell Proliferation (MTT) Assay

This MTT assay is used to determine the IC₅₀ concentration of Tribulus terrestris extract on MCF-7 cells [43]. For MTT assay, cells were placed in 96-well plates at the density of 10000 cells/well and were incubated for 24 hours at 37°c in 5% CO₂ for attachment of cells. After 24 hours various concentrations (50,100, 150, 200, 250, 300 µg/ml) of Tribulus terrestris were added to the cells and incubated for 24 hrs at 37°C. After incubation, the medium was replaced with 10 µl µl of MTT (5 mg/ml) dye in serum free medium was added per well and wrapped with aluminium foil and incubated for further 4 hours dark at 37°C. Then, 100 µl DMSO was added to the wells to solubilize the formazan crystals. The absorbance was measured at 570 nm. The percentage of cell inhibition was determined by following formula

2.5 Cell Morphological Studies by Phase Contrast Microscope

Morphological changes in the cancer cells before and after *Tribulus terrestris* extract treatment can be studied with the help of phase contrast microscope. At the end of the experiment the cells were taken and observed under an inverted light microscope with 20x magnification.

2.6 Statistical Analysis

All the data from the results were analyzed by ttest and depicted as mean \pm SD. The results were statistically analysed using one way ANOVA in SPSS software. The statistical significance was at p<0.05.

3. RESULTS AND DISCUSSION

Tribulus terrestris is one of the plants which is used for a long time in treating various diseases [44]. The plant is known for its anticancer, antidiabetic. antispasmodic, antifungal. antibacterial, anticariogenic and many more activities which make this a vital medicine for many ailments [45]. Saponins present in the plant are found to be responsible for antifungal and antibacterial [46]. The reason for choosing this plant is that it has a role in reducing hormonal imbalance and increasing reproductive function. Dysregulated proliferation of cells is seen in various types of cancer [47]. In many countries, cancers are evolving as a reason for malignancy associated with death [48]. Many drugs are being discovered for potent protective effects against cancer cells [49]. In a previous study by Arumugam et al., it was discussed that cell cycle arrest by compounds can apoptitize cancerous cells and end proliferation of cells [50]. In the present study, the cell viability assay showed that, cell growth inhibition was observed in dose dependent manner. The ethanolic extract Tribulus terrestris treatment significantly of reduces the breast cancer cell viability. At 40 µg/ml concentration of the extract arrests 50% of the human breast cancer cells proliferation. And also, Tribulus terrestris extract (40 µg/ml) treatment significantly induces the apoptosis, was evidenced in phase contrast which microscopic analysis. The cell morphological changes observed in Tribulus terrestris extract (40 µg/ml) treated reast cancer cells at 24hrs time point.

In the study by Apurva et al., the extract of the *Tribulus terrestris* plant seems to increase the caspase 3 activity of the human breast cancer cell line [51]. In the study by Masoud et al., the herb is also found to have anticancer activity against prostate and colon cancer cell lines [52]. It was inferred that the aqueous extract of the plant had a high cytotoxic effect and it can be

suggested for marketing in the study by Claudio et al. [53]. In the study by Farooq et al., it was inferred that saponins present were responsible for the anti tumor and anti proliferative function of the plant [54]. In the present study, it is clear that the anti cancer activity of the ethanolic extract of *Tribulus terrestris* plant is completely concentration dependent. At different concentrations, the activity of the extract differs. The limitation is that it is an *in vitro* study. Further experimental validation is required for establishment as an anticancer treatment against human breast cancer cells [24, 55-68].

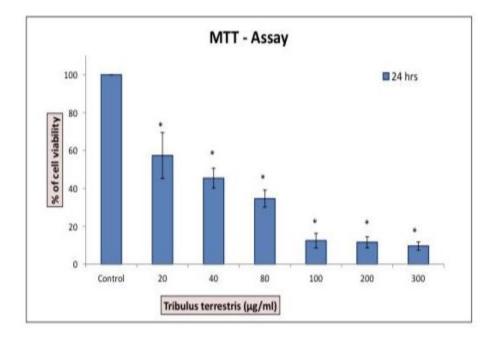
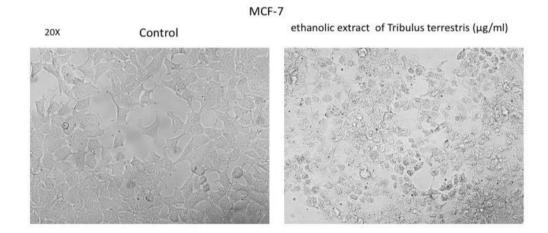
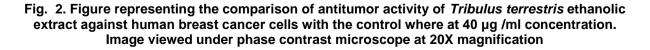


Fig. 1. Bar graph representing the antitumor activity of *Tribulus terrestris* ethanolic extract via MTT assay at a regular interval of 24 hours. X axis represents the different concentrations of *Tribulus terrestris* ethanolic extract in (microgram/ ml) while Y axis represents the percentage of cell viability (in numbers). Data are shown as means \pm SD (n = 3). * compared with the control-blank group, p < 0.001





4. CONCLUSION

From the above results, the *Tribulus terrestris* extracts were cytotoxic to the human breast cancer cell line MCF-7 at this dose dependent concentration. However more research is needed to understand the mechanisms of cytotoxicity and this study provides scope for future studies.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Sun YS, Zhao Z, Yang ZN, Xu F, Lu HJ, Zhu ZY, et al. Risk Factors and Preventions of Breast Cancer. Int J Biol Sci. 2017 Nov 1;13(11):1387–97.
- Akram M, Iqbal M, Daniyal M, Khan AU. Awareness and current knowledge of breast cancer. Biological Research. 2017;50.

Available:http://dx.doi.org/10.1186/s40659-017-0140-9

- McKinney SM, Sieniek M, Godbole V, Godwin J, Antropova N, Ashrafian H, et al. Addendum: International evaluation of an AI system for breast cancer screening. Nature. 2020 Oct;586(7829):E19.
- 4. Sharma GN, Dave R, Sanadya J, Sharma P, Sharma KK. Various types and management of breast cancer: an overview. J Adv Pharm Technol Res. 2010;1(2):109-126.
- 5. Zhu W, Du Y, Meng H, Dong Y, Li L. A review of traditional pharmacological uses, phytochemistry, and pharmacological activities of *Tribulus terrestris*. Chem Cent J. 2017 Jul 11;11(1):60.
- Semerdjieva IB, Zheljazkov VD. Chemical Constituents, Biological Properties, and Uses of *Tribulus terrestris*: A Review. Natural Product Communications. 2019; 14:1934578-1986839. Available:http://dx.doi.org/10.1177/193457 8x19868394

- Journal BS, Baghdad Science Journal. Effect of Aqueous and Ethanolic Extracts of *Tribulus terrestris*, Phoenix dactylifera and Nasturtium officinale Mixture on Some Reproductive Parameters in Male Mice . Baghdad Science Journal. 2012;9:640–50. Available:http://dx.doi.org/10.21123/bsj.9.4 .640-650
- Chhatre S, Nesari T, Somani G, Kanchan D, Sathaye S. Phytopharmacological overview of *Tribulus terrestris*. Pharmacogn Rev. 2014 Jan;8(15):45–51.
- Rajeshkumar S, Kumar SV, Ramaiah A, Agarwal H, Lakshmi T, Roopan SM. Biosynthesis of zinc oxide nanoparticles usingMangifera indica leaves and evaluation of their antioxidant and cytotoxic properties in lung cancer (A549) cells. Enzyme Microb Technol. 2018 Oct;117:91–5.
- Vairavel M, Devaraj E, Shanmugam R. An eco-friendly synthesis of Enterococcus sp.-mediated gold nanoparticle induces cytotoxicity in human colorectal cancer cells. Environ Sci Pollut Res. 2020 Mar 1;27(8):8166–75.
- Gomathi M, Prakasam A, Rajkumar PV, 11. Rajeshkumar S, Chandrasekaran R. Anbarasan PM. Green synthesis of silver nanoparticles using Gymnema sylvestre leaf extract and evaluation of its antibacterial activity. South African Journal of Chemical Engineering. 2020;32:1-4. Available:http://dx.doi.org/10.1016/j.sajce.2 019.11.005
- 12. Rajasekaran S, Damodharan D, Gopal K, Rajesh Kumar B, De Poures MV. Collective influence of 1-decanol addition, injection pressure and EGR on diesel engine characteristics fueled with diesel/LDPE oil blends. Fuel. 2020 Oct 1;277:118166.
- Santhoshkumar J, Sowmya B, Venkat Kumar S, Rajeshkumar S. Toxicology evaluation and antidermatophytic activity of silver nanoparticles synthesized using leaf extract of Passiflora caerulea. S Afr J Chem Eng. 2019 Jul;29:17–23.
- Raj RK, DE, SR. β-Sitosterol-assisted silver nanoparticles activates Nrf2 and triggers mitochondrial apoptosis via oxidative stress in human hepatocellular cancer cell line. J Biomed Mater Res A. 2020 Sep;108(9):1899–908.
- 15. Saravanan M, Arokiyaraj S, Lakshmi T, Pugazhendhi A. Synthesis of silver nanoparticles from Phenerochaete

chrysosporium (MTCC-787) and their antibacterial activity against human pathogenic bacteria. Microb Pathog. 2018 Apr;117:68–72.

- 16. Gheena S, Ezhilarasan D. Syringic acid triggers reactive oxygen species-mediated cytotoxicity in HepG2 cells. Hum Exp Toxicol. 2019 Jun 1;38(6):694–702.
- Ezhilarasan D, Sokal E, Najimi M. Hepatic fibrosis: It is time to go with hepatic stellate cell-specific therapeutic targets. Hepatobiliary Pancreat Dis Int. 2018 Jun;17(3):192–7.
- Ezhilarasan D. Oxidative stress is bane in chronic liver diseases: Clinical and experimental perspective. Arab J Gastroenterol. 2018 Jun;19(2):56–64.
- 19. Gomathi AC, Xavier Rajarathinam SR, Mohammed Sadiq A, Rajeshkumar S. Anticancer activity of silver nanoparticles synthesized using aqueous fruit shell extract of Tamarindus indica on MCF-7 human breast cancer cell line. J Drug Deliv Sci Technol. 2020 Feb 1;55:101376.
- Dua K, Wadhwa R, Singhvi G, Rapalli V, Shukla SD, Shastri MD, et al. The potential of siRNA based drug delivery in respiratory disorders: Recent advances and progress. Drug Dev Res. 2019 Sep;80(6):714–30.
- Ramesh A, Varghese S, Jayakumar ND, Malaiappan S. Comparative estimation of sulfiredoxin levels between chronic periodontitis and healthy patients - A casecontrol study. J Periodontol. 2018 Oct;89(10):1241–8.
- 22. Arumugam P, George R, Jayaseelan VP. Aberrations of m6A regulators are associated with tumorigenesis and metastasis in head and neck squamous cell carcinoma. Arch Oral Biol. 2021 Feb;122:105030.
- 23. Joseph B, Prasanth CS. Is photodynamic therapy a viable antiviral weapon against COVID-19 in dentistry? Oral Surg Oral Med Oral Pathol Oral Radiol. 2021 Jul;132(1):118–9.
- Ezhilarasan D, Apoorva VS, Ashok VN. Syzygium cumini extract induced reactive oxygen species-mediated apoptosis in human oral squamous carcinoma cells. J Oral Pathol Med . 2019 Feb [cited 2021 Sep 15];48(2). Available:https://pubmed.ncbi.nlm.nih.gov/

30451321/ 25. Duraisamy R, Krishnan CS, Ramasubramanian H, Sampathkumar J, Mariappan S, Navarasampatti Sivaprakasam A. Compatibility of Nonoriginal Abutments With Implants: Evaluation of Microgap at the Implant-Abutment Interface, With Original and Nonoriginal Abutments. Implant Dent. 2019 Jun;28(3):289–95.

- Rajeshkumar S, Ezhilarasan D, Puyathron N, Lakshmi T. Role of supermagnetic nanoparticles in Alzheimer disease. In: Nanobiotechnology in Neurodegenerative Diseases. Cham: Springer International Publishing; 2019: 225–40.
- Rajeshkumar S, Lakshmi T, Tharani M, Sivaperumal P. Green synthesis of gold nanoparticles using pomegranate peel extract and its antioxidant and anticancer activity against liver cancer cell line. Alınteri zirai bilim derg. 2020 Nov 27;35(2):164–9.
- Rajeshkumar S, Tharani M, Sivaperumal P, Lakshmi T. Synthesis of Antimicrobial Silver Nanoparticles by Using Flower of Calotropis Gigantea. Journal of Complementary Medicine Research. 2020;11(5):8–16.
- 29. Lakshmi T, Ezhilarasan D, Nagaich U, Vijayaragavan R. Acacia catechu Ethanolic Seed Extract Triggers Apoptosis of SCC-25 Cells. Pharmacogn Mag . 2017 Oct [cited 2021 Aug 31];13(Suppl 3). Available:
- https://pubmed.ncbi.nlm.nih.gov/29142391/
 30. Phyto-assisted synthesis of zinc oxide nanoparticles using Cassia alata and its antibacterial activity against Escherichia coli. Biochemistry and Biophysics Reports. 2019 Mar 1;17:208–11.
- Rajeshkumar S, Sivaperumal P, Tharani M, Lakshmi T. Green Synthesis of Zinc Oxide Nanoparticles by Cardiospermum -. Journal of Complementary Medicine Research. 2020;11(5):128–36.
- Rajeshkumar S, Tharani M, Sivaperumal P, Lakshmi T. Green Synthesis of Selenium Nanoparticles Using Black Tea (Camellia Sinensis) And Its Antioxidant and Antimicrobial Activity. Journal of Complementary Medicine Research. 2020;11(5):75–82.
- R. Jagadheeswari RJ, T. Lakshmi TL, Balusamy SR, David S, Kumar SR. Biosynthesis of silver nanoparticles using *Withania somnifer*a (L.) Dunal extract and its antibacterial activity against food pathogens. Ann Phytomed. 2020 Jun; 9(1).

Available:http://www.ukaazpublications.co m/publications/?smd_process_download= 1&download_id=9526

- Molecular docking analysis of compounds from Lycopersicon esculentum with the insulin receptor to combat type 2 diabetes . [cited 2021 Aug 31]. Available:http://www.bioinformation.net/01 6/97320630016748.htm
- 35. Anticancer effects and lysosomal acidification in A549 cells by Astaxanthin from Haematococcus lacustris . [cited 2021 Aug 31]. Available:http://www.bioinformation.net/01

6/97320630016965.htm

- Akshayaa L, Lakshmi, Thangavelu, Devaraj, Ezhilarasan, Roy, Anitha, Raghunandhakumar, S, Sivaperumal P, David, Sheba, Dua, Kamal, Chellappan, Dinesh Kumar. Data on known anti-virals in combating CoVid-19. Bioinformation. 2020;878–878.
- 37. Rajeshkumar S, Agarwal H, Sivaperumal Shanmuqam Ρ. VK. Lakshmi Τ. Antimicrobial. anti-inflammatory and anticancer potential of Microbes mediated zinc oxide nanoparticles. Journal of Complementary Medicine Research. 2020;11(5):41-8.
- Thangavelu L, Balusamy SR, Shanmugam R, Sivanesan S, Devaraj E, Rajagopalan V, et al. Evaluation of the sub-acute toxicity of Acacia catechu Willd seed extract in a Wistar albino rat model. Regul Toxicol Pharmacol . 2020 Jun [cited 2021 Aug 31];113.

Available:https://pubmed.ncbi.nlm.nih.gov/ 32169672/

- Cytotoxic potentials of silibinin assisted silver nanoparticles on human colorectal HT-29 cancer cells . [Cited 2021 Aug 31]. Available:http://www.bioinformation.net/01 6/97320630016817.htm
- 40. Shaker Ardakani L, Surendar A, Thangavelu L, Mandal T. Silver nanoparticles (Ag NPs) as catalyst in chemical reactions. Synth Commun. 2021 Mar 8;1–21.
- 41. Hashim IM, Ghazi IF, Kuzichkin OR, Shakirova IA, Surendar A, Thangavelu L, et al. Effects of Primary Stored Energy on Relaxation Behavior of High Entropy Bulk Metallic Glasses Under Compressive Elastostatic Loading. Trans Indian Inst Met. 2021 Mar 14;74(6):1295– 301.

42. Krishnan V, Lakshmi T. Bioglass: A novel biocompatible innovation. J Adv Pharm Technol Res. 2013 Apr [Cited 2021 Aug 31];4(2). Available:

https://pubmed.ncbi.nlm.nih.gov/23833747/
43. Koka P, Mundre RS, Rangarajan R, Chandramohan Y, Subramanian RK, Dhanasekaran A. Uncoupling Warburg effect and stemness in CD133 cancer stem cells from Saos-2 (osteosarcoma) cell line under hypoxia. Mol Biol Rep. 2018

- Dec;45(6):1653–62.
 44. Zhang J-D, Xu Z, Cao Y-B, Chen H-S, Yan L, An M-M, et al. Antifungal activities and action mechanisms of compounds from *Tribulus terrestris* L. J Ethnopharmacol. 2006 Jan 3;103(1):76–84.
- 45. Sahani K, Thakur D, Hemalatha KPJ. Phytochemical analysis and antioxidant activity of endophytic fungi Curvularia aeria MTCC 12847 isolated from *Tribulus terrestris* L. leaf. Available: http://dx.doi.org/10.21203/rs.2.16923/v1
- Yuan W-H. Two Furostanol Saponins from the Fruits of *Tribulus terrestris*. Vol. 6, Chinese Journal of Natural Medicines. 2008:172–5. Available:http://dx.doi.org/10.3724/sp.j.100 9.2008.00172
- 47. Raghunandhakumar S, Paramasivam A, Senthilraja S, Naveenkumar C, Asokkumar S, Binuclara J, et al. Thymoquinone inhibits cell proliferation through regulation of G1/S phase cell cycle transition in Nnitrosodiethylamine-induced experimental rat hepatocellular carcinoma. Toxicol Lett. 2013 Oct 23;223(1):60–72.
- 48. Asokkumar S. Naveenkumar C. Raghunandhakumar S. S, Kamarai Anandakumar P, Jagan S. et al. Antiproliferative and antioxidant potential of beta-ionone against benzo(a)pyreneinduced lung carcinogenesis in Swiss albino mice. Mol Cell Biochem. 2012 Apr;363(1-2):335-45.
- 49. Anandakumar P, Kamaraj S, Jagan S, Ramakrishnan G, Asokkumar S, Naveenkumar C, et al. Capsaicin inhibits benzo(a)pyrene-induced lung carcinogenesis in an in vivo mouse model. Inflamm Res. 2012 Nov;61(11): 1169–75.
- 50. Paramasivam A, Raghunandhakumar S, Priyadharsini JV, Jayaraman G. In Vitro Anti-Neuroblastoma Activity of Thymoquinone Against Neuro-2a Cells via

Cell-cycle Arrest. Asian Pac J Cancer Prev. 2015;16(18):8313–9.

- Patel A, Soni A, Siddiqi NJ, Sharma P. An insight into the anticancer mechanism of *Tribulus terrestris* extracts on human breast cancer cells . 2019;9(3)Biotech. Available:http://dx.doi.org/10.1007/s13205-019-1585-z
- 52. Pourali M, Yaghoobi MM, Sormaghi MHS. Cytotoxic, Anti-Proliferative and Apoptotic Effects of *Tribulus terrestris* L. Fruit Extract on Human Prostate Cancer Lncap and Colon Cancer HT-29 Cell Lines. Vol. Inpress, Jundishapur Journal of Natural Pharmaceutical Products. 2016. Available: http://dx.doi.org/10.17795/jjnpp-33561
- Filho CCO, Kampke EH, Vargas TS, Salustriano NA, Scherer R, Fronza M, et al. In vitro cytotoxic activity of five commercial samples of *Tribulus terrestris* Linn in Espírito Santo (Brazil) . Vol. 53, Brazilian Journal of Pharmaceutical Sciences. 2018. Available: http://dx.doi.org/10.1590/s2175-

Available: http://dx.doi.org/10.1590/s2175-97902017000400262

- 54. Farooq SA, Farook TT, Al-Rawahy SH. Bioactive compounds from Tribulus terrestris L.(zygophyllaceae). Natural Products and Their Active Compounds on Disease Prevention: Nova Science Publishers: Hauppauge, NY, USA. 2012;245-68.
- 55. Danda AK, Krishna TM, Narayanan V, Siddareddi A. Influence of primary and secondary closure of surgical wound after impacted mandibular third molar removal on postoperative pain and swelling--a comparative and split mouth study. J Oral Maxillofac Surg . 2010 Feb [cited 2021 Sep 15];68(2). Available:https://pubmed.ncbi.nlm.nih.gov/

Available:https://pubmed.ncbi.nlm.nih.gov/ 20116700/

56. Ramadurai N, Gurunathan D, Samuel AV, Subramanian E, Rodrigues SJL. Effectiveness of 2% Articaine as an anesthetic agent in children: randomized controlled trial. Clin Oral Investig . 2019 Sep [cited 2021 Sep 15];23(9). Available:

https://pubmed.ncbi.nlm.nih.gov/30552590/ 57. Sathivel A, Raghavendran HR, Srinivasan

P, Devaki T. Anti-peroxidative and antihyperlipidemic nature of Ulva lactuca crude polysaccharide on D-galactosamine induced hepatitis in rats. Food Chem Toxicol . 2008 Oct [cited 2021 Sep 15];46(10). Available:https://pubmed.ncbi.nlm.nih.gov/ 18706469/

58. Panda S, Doraiswamy J, Malaiappan S, Varghese SS, Del Fabbro M. Additive effect of autologous platelet concentrates in treatment of intrabony defects: a systematic review and meta-analysis. J Investig Clin Dent . 2016 Feb [cited 2021 Sep 15];7(1).

Available:https://pubmed.ncbi.nlm.nih.gov/ 25048153/

- Neelakantan P, Varughese AA, Sharma S, Subbarao CV, Zehnder M, De-Deus G. Continuous chelation irrigation improves the adhesion of epoxy resin-based root canal sealer to root dentine. Int Endod J . 2012 Dec [cited 2021 Sep 15];45(12). Available:https://pubmed.ncbi.nlm.nih.gov/ 22612994/
- Govindaraju L, Neelakantan P, Gutmann JL. Effect of root canal irrigating solutions on the compressive strength of tricalcium silicate cements. Clin Oral Investig . 2017 Mar [cited 2021 Sep 15];21(2). Available:https://pubmed.ncbi.nlm.nih.gov/ 27469101/
- 61. Sekhar CH, Narayanan V, Baig MF. Role of antimicrobials in third molar surgery: prospective, double blind,randomized, placebo-controlled clinical study. Br J Oral Maxillofac Surg . 2001 Apr [cited 2021 Sep 15];39(2).

Available:https://pubmed.ncbi.nlm.nih.gov/ 11286448/

- DeSouza SI, Rashmi MR, Vasanthi AP, Joseph SM, Rodrigues R. Mobile phones: the next step towards healthcare delivery in rural India? PLoS One . 2014 Aug 18 [cited 2021 Sep 15];9(8). Available:https://pubmed.ncbi.nlm.nih.gov/ 25133610/
- Nasim I, Neelakantan P, Sujeer R, Subbarao CV. Color stability of microfilled, microhybrid and nanocomposite resins--an in vitro study. J Dent . 2010 [Cited 2021 Sep 15];38 Suppl 2. Available:https://pubmed.ncbi.nlm.nih.gov/ 20553993/
- 64. Danda AK, Muthusekhar MR, Narayanan V, Baig MF, Siddareddi A. Open versus closed treatment of unilateral subcondylar and condylar neck fractures: a prospective, randomized clinical study. J Oral Maxillofac Surg . 2010 Jun

[Cited 2021 Sep 15];68(6).

Available:https://pubmed.ncbi.nlm.nih.gov/20303209/

Kamalli et al.; JPRI, 33(62A): 387-395, 2021; Article no.JPRI.77808

- 65. Molecular structure and vibrational spectra of 2.6-bis(benzvlidene)cvclohexanone: A densitv functional theoretical studv. Spectrochim Acta А Mol Biomol Spectrosc. Jan 2011 1;78(1):113-21.
- Putchala MC, Ramani P, Herald J. Sherlin, Premkumar P, Natesan A. Ascorbic acid and its pro-oxidant activity as a therapy for tumours of oral cavity – A systematic review. Archives of Oral Biology. 2013;58:P 563–74. Available:http://dx.doi.org/10.1016/j.archor albio.2013.01.016
- 67. Neelakantan P, Grotra D, Sharma S. Retreatability of 2 mineral trioxide aggregate-based root canal sealers: a cone-beam computed tomography analysis. J Endod. 2013;39(7):893–6.
- Suresh P, Marimuthu K, Ranganathan S, Rajmohan T. Optimization of machining parameters in turning of Al-SiC-Gr hybrid metal matrix composites using grey-fuzzy algorithm. Transactions of Nonferrous Metals Society of China. 2014;24: 2805–14. Available: http://dx.doi.org/10.1016/s1003-6326(14)63412-9

© 2021 Kamalli et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/77808