

Asian Journal of Research in Medical and Pharmaceutical Sciences

7(1): 1-9, 2019; Article no.AJRIMPS.48844 ISSN: 2457-0745

Comparison of the Hyperglycemic Control of *M. oleifera* Leaves Aqueous Extract and Glibenclamide Tablets in Alloxan Monohydrate Induced Diabetic Rats

Josephine N. Kasolo^{1*}, Agnes Namaganda¹, Joshua Nfambi¹, Ivan Kimuli¹, Haruna Muwonge¹ and Isaac Okullo²

> ¹Department of Physiology, Makerere College of Health Sciences, Uganda. ²Department of Dental Surgery, Makerere College of Health Sciences, Uganda.

Authors' contributions

This work was carried out in collaboration among all authors. Author JNK designed the study, performed the statistical analysis, wrote the protocol and wrote the first draft of the manuscript. Author AN participated in the data collection. Authors JN and IK managed the literature review. Author HM managed the analysis of the study. Author IO was the overall mentor of the study. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/AJRIMPS/2019/v7i130112 <u>Editor(s):</u> (1) Dr. S. Prabhu, Department of Biotechnology, Sri Venkateswara College of Engineering, Sriperumbudur, India. <u>Reviewers:</u> (1) Márcio Rodrigues, University of Beira Interior, Portugal. (2) Rupali Sengupta, SNDT Women's University, India. Complete Peer review History: <u>http://www.sdiarticle3.com/review-history/48844</u>

Original Research Article

Received 28 January 2019 Accepted 23 April 2019 Published 30 April 2019

ABSTRACT

Introduction: Diabetes being one of the commonest non-communicable diseases worldwide has no cure. The available hypoglycemic drugs are costly, and have associated long-term side effects. *M. oleifera* leaves are used in many countries in Africa and Asia to treat diabetes. The study compared the hyperglycemic control of *M. oleifera* leaves aqueous extract and Glibenclamide tablet in alloxan monohydrate induced diabetic rats.

Methods: Twenty-four female Wistar albino rats, made diabetic using alloxan monohydrate, received either *M. oleifera* extract, glibenclamide or distilled water were delivered intragastric. The mean body weight and mean fasting blood sugar were measured over a period of 28 days.

*Corresponding author: E-mail: jkasolo@chs.mak.ac.ug, josephinekasolo57@gmail.com;

Results: Rats that received distilled water had a mean fasting blood sugar of 329.3±44.9 mg/dl at the beginning, which increased to 448.0±189.9 mg/dl on day 14; all the rats were dead by day 21. The rats that received *M. oleifera* had blood sugar 443.4±134.7 mg/dl at the beginning, dropped to 166.5±162.79 mg/dl by day 14, and to 88.7±41.0 mg/dl by day 28. Rats that received glibenclamide had blood sugar 517.6±139.3mg/dl at the beginning, dropped to 209.0±201.9 mg/dl on day 14, and to 89.7±42.85 mg/dl on day 28. The blood sugar of the *M. oleifera* and glibenclamide groups reached normal level by day 21 and remained within the normal range up to day 28. *Conclusion: Moringa oleifera* leaves aqueous extract has similar pattern to glibenclamide tablet in causing hypoglycemia to alloxan monohydrate induced diabetic rats.

Keywords: M. oleifera; alloxan induced diabetes; glibenclamide; hypoglycemia; Wistar rats.

1. INTRODUCTION

Diabetes mellitus is a serious disease with no cure, which is costly and is becoming increasingly common, especially in developing countries and disadvantaged minorities. It continues to be a global public health problem with affected individuals rising from 108 million in 1980 to 422 million in 2014, and with middle- and low-income countries most affected [1]. In Uganda, diabetes mellitus is at 2.8% prevalence [2], which closely compares with other countries in East Africa.

Diabetes mellitus (DM) is generally associated with metabolic disorders plus inflammation and oxidative stress. The disease is characterized by hyperglycemia and hyperlipidemia, which result from disturbances in carbohydrate, protein, and lipid metabolism. Type1 diabetes (T1D), also termed juvenile-onset or insulin-dependent diabetes, is an autoimmune disease and a metabolic disorder characterized by T-cellmediated destruction of pancreatic beta (β) cells, resulting in insulin deficiency and hyperglycaemia. Type 2 diabetes (T2D), (noninsulin-dependent) DM which occurs in adults, is caused by insulin resistance coupled with a failure of the β cell to compensate.

Chronic inflammation has been indicated as a risk factor for the development of type 2 diabetes with increasing evidence pointing toward a role of pro-inflammatory cytokines such as C-reactive protein (CRP), interleukin (IL)-6, and tumor necrosis factor (TNF α) in the pathogenesis of insulin resistance and type 2 diabetes [3,4,5].

Research evidence indicates that several trace elements are essential for normal glucose homeostasis that includes: chromium, potassium, calcium, magnesium, copper, manganese and zinc [6]. Vitamin B1 as well as vitamins B6 and B12 support nervous system functions and helps prevent diabetic neuropathies. Targeted consumption of micronutrients can help to improve metabolic control, optimize treatment and reduce the risk of developing diabetic complications. As coenzymes, the B vitamins play a central role in carbohydrate, protein and lipid metabolism. Studies indicate that the majority of type 1 and type 2 diabetics have inadequate supplies of vitamin B1 and impaired thiamine metabolism [7]. Lack of folic acid and/or vitamin B12 leads to impaired metabolism of the amino acid methionine and is frequently accompanied by elevated plasma homocysteine concentration [8]. In diabetics, increased oxidative stress may be a result of decreased plasma concentration of the antioxidant vitamins C and E, coupled with the reduced postprandial intracellular ratio of ascorbic acid to its oxidized form (dehydroascorbic acid) [9].

Recent studies indicate that an inadequate supply of vitamin D could be involved in the onset of numerous chronic diseases like diabetes mellitus types 1 and 2 [10, 11]. On the other hand, lack of vitamin D represents a risk factor for type2 diabetes and metabolic syndrome, since it increases insulin resistance and reduces insulin secretion from pancreatic beta cells [12]. There is evidence that vitamin D can help to prevent the destruction of insulin-producing pancreatic beta cells and thus combat the onset of type 1 diabetes [13]. The effects of vitamin D are assumed to be due primarily to the immunomodulatory action of the vitamin via Thelper cells and to the reduction of proinflammatory cytokines.

The therapeutic management of diabetes without any side effects remains a challenge. However, there is a growing interest in evaluating herbal remedies, which are seen to be less toxic with negligible side effects [14]. One such a plant is *Moringa oleifera* Lam. *M. oleifera* leaves have for long time been used in folk medicine to treat diabetes in different communities of the world including Uganda [15, 16,17]. *M. oleifera* is a rapid growing tree, native to the Sub-Himalayan tracts of India, Pakistan, Bangladesh and Afghanistan. It was utilized by the ancient Romans, Greeks, Egyptians and Indians to treat several ailments [18].

The *M. oleifera* plant is drought tolerant and is known to thrive best in tropical conditions. The plant also tolerates different soil types, and boasts as one of the few medicinal plants which is well documented. The scientific classification of *Moringa oleifera* shows that it belongs to the Kingdom: Plantae; Division: Magnoliphyta; Class: Magnoliopsida; Order: Brassicales; Family: Moringaceae; Genus: Moringa; Species: *Moringa oleifera. Moringa oleifera* is the most widely cultivated variety of the genus Moringa in Asia and Africa [19, 0].

1.1 Hypoglycemic Potential of *M. Oleifera* Leaves

Moringa oleifera leaf extract possess potent hypoglycemic effects through the normalization of elevated hepatic pyruvate carboxylase enzyme and regeneration of damaged hepatocytes and pancreatic cells in rats, and also via its antioxidant properties on the liver and pancreas, plus an increase in β -cell mass and insulin production by the β -cells [21].

The phytochemicals that exist in M. oleifera are capable of acting on animal cells and tissues to inhibit membrane bound enzymes, which affect DNA formation and destroy cell membranes [22, 23, 24]. The leaves have antioxidants which can combine with reactive oxygen species to prevent cell damage that is believed to occur in diabetes mellitus [25,26,27]. Additionally, the methanol extract of M. oleifera was found to have immunomodulatory activity in rats, which could be useful in treating type 1 diabetes [28]. The therapeutic potential and medicinal properties of M. oleifera leaves has been evaluated in a number of studies using animal models and they have been proved to have hypoglycemic activities [29,30]. Extracts of M. oleifera leaves also have anti-inflammatory activities which are reported to contribute to the hypogylycemic activities [31,32,33].

Safety evaluation of *Moringa oleifera* leaves done by researchers found them safe for human and animal consumption by WHO standards [34,35,36,37,38]. There are few studies that have compared the hyperglycemic control of herbal medicines with orthodox medicines on the market [39]. The aim of this was to compare the hyperglycemic control of *Moringa oleifera* leaves aqueous extract and glibenclamide tablets on alloxan monohydrate induced hyperglycemia in Wistar albino rats. Ethical approval was obtained from the Institution Review Board (IRB) of the Makerere University College of Health Sciences, School of Biomedical Sciences, SBS-HDREC 565. The animals were treated humanely according to international standard OECD guidelines (2001).

2. MATERIALS AND METHODS

2.1 Plant Handling and Extraction

Fresh mature green leaves of *Moringa oleifera* were collected from Wakiso district, Uganda's central region, growing on the hillside loam soil, harvested during the rainy season between 9.00 and 11.00 a.m. Plant species and family were confirmed by a Makerere University plant taxonomist, and a specimen voucher number (41302) deposited at the Makerere University herbarium. It was air-dried in a shade for about 3 weeks until constant weight was attained, away from direct sun shine to protect the active compounds. The leaves were pulverized into coarse powder using a mortar and pestle to ease the extraction.

2.2 Moringa oleifera Phytochemical Extraction

Serial extraction which followed the established method starting with ether, then ethanol and lastly water was done [40,41]. Briefly, one liter of ether was mixed with 500gm of M. oleifera leaf powder and shaken at intervals for two days. The mixture was decanted and filtered. The residue was air-dried for 3 days and 700 ml of ethanol (98% V/V) added, and left to evaporate until it dried. The dry residue was then soaked in 700 ml of warm water at 40°C to facilitate the extraction. The ether and ethanol solvents were recovered using a rotary evaporator (BUCHI Rotavapor R-205) while the water extract was freeze dried into powder. The powder was dissolved in distilled water to make a stock solution from which the rats were dosed.

2.3 Study Animals

This study used 24 female Wistar rats aged 8-10 weeks, reared in the Makerere University, College of Veterinary Medicine, Animal

Kasolo et al.; AJRIMPS, 7(1): 1-9, 2019; Article no.AJRIMPS.48844

Resources and Biosafety's animal house. The experiment was carried out in the animal house at the department of Physiology, Makerere University College of Health Sciences. The animals received 12 hrs of light and 12 hrs of darkness, fed on commercial rat pellets and allowed to take food and tap water *ad libitum*. The rat housing was kept at room temperature. The rats were of normal body temperature, active and feeding well; and weighed 90-110 gm each. Pregnant or Nursing rats were excluded from the study.

2.4 Induction of Hyperglycemia

Alloxan monohydrate (Sigma, St. Louis, MI, USA) was used to induce hyperglycemia in the rats. Alloxan monohydrate was dissolved in 0.9% normal saline and injected intraperitoneally in a single dose of 100 mg/kg body weight to overnight-fasted rats [34]. Interventions were introduced when the rats showed fasting blood glucose levels >250 mg/dL, as well as a reduction in body weight with signs of polyphagia, polyuria, and polydipsia.

2.5 Dosing of Animals

The animals were randomly allocated to 3 groups of 8 rats each. The rats in each group were made diabetic using alloxan monohydrate. Each group received the intervention intragastric once a day for 28 days. Food was withdrawn from the rats at 10.00p.m, but they were allowed to take tap water *ad libitum*. Food was reintroduced after weighing and measuring blood sugar. The rats were allocated to different groups as follows:

Group I: Diabetic rats that received 1 ml distilled water once daily for 28 days (negative control). Group II. Diabetic rats that received 500 mg/kg of *Moringa oleifera* aqueous extract once daily for 28 days. Group III: Diabetic rats received 0.04 mg/kg tablet Glibenclamide (positive control) once

daily for 28 days. On a weekly basis, body weight and fasting

blood sugar for each rat was measured between 8.00 and 9.00 a.m, using "On Call plus Blood Glucose Meter" glucometer purchased from Acon Laboratories, Inc. 10125 Mesa Rim Road, San Diego, CA92121, USA from an ear lobe prick.

2.6 Data Analysis

The data was analyzed using Prism 7 (GraphPad) software (SanDiego, CA, USA)

where the means and standard deviations were compared. The data from each intervention group were compared on days 7, 14, 21 and 28 against the results of day 1 using ordinary one way ANOVA with Dunnets adjustment for multiple comparisons. The level of significance was fixed at $p \le 0.05$.

3. RESULTS

Fig. 1 shows that the mean body weight of alloxan diabetic rats during the 28 days of treatment. The rats that received 1 ml distilled water all died by the third week of the study, while those that received 500 mg/kg M. oleifera leaves aqueous extract and those that received 0.4 mg/kg glibenclamide tablets survived up to day 28. Generally, the mean body weight of the diabetic rats that received М. oleifera aqueous extract and glibenclamide increased although the increase slightly, was not significant. In contrast, the mean body weight of the diabetic rats that received 1 ml distilled water had reduced by day 14, although the decrease was not significant.

Fig. 2, shows that all the animals had blood sugar of more than 300mg/dl on day one before Interventions. In the group that received distilled water, fasting blood sugar significantly rose during the intervention period, leading to all rats dying before the end of the third week. However, the fasting blood sugar levels of the animals that received M. oleifera leaves aqueous extract and glibenclamide tablets dropped significantly between day 1 and day 7, and then again between day 14 and day 21. Between day 7 and day 14, the reduction in fasting blood glucose was not significant in the rats that received an aqueous extract of М. oleifera leaves, and glibenclamide. By day 21, the fasting blood sugar levels in both groups of rats treated with an aqueous extract of M. oral hypoglycemic oleifera leaves, and agent glibenclamide had returned to normal, with mean blood glucose levels of 92.7+28.38 mg/dl, and 74.0+10.15 mg/dl in the aqueous M. oleifera and glibenclamide treatment groups respectively.

During the 4th week of the study (days 22 to 28) the change in fasting blood sugar in the rats was minimal and it remained in the normal range, with the mean<u>+</u>SD of the *M. oleifera* extract and glibenclamide treatment groups being 88.7 ± 41.00 mg/dl, and 89.7 ± 42.85 mg/dl respectively. In general the graphs of *M. oleifera*

extract and tablet glibenclamide interventions had a similar pattern.

4. DISCUSSION

The hypoglycemic effect of *M. oleifera* leaves aqueous extract and glibenclamide tablets on alloxan monohydrate induced diabetic rats were

investigated. Study results show that the aqueous extract of *M. oleifera* leaves grown on Ugandan soil have hypoglycemic effects in Wister albino rats. They also show that there was insignificant increase in the mean body weight for *M. oleifera* extract and the glibenclamide group which is an indicator of diabetes control.

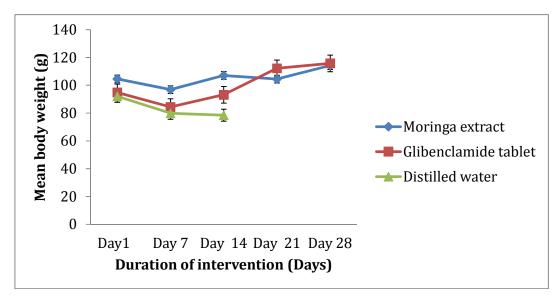


Fig. 1. Mean body weight (g) against days during intervention for *M. oleifera* leaves aqueous extract, glibenclamide tablets and distilled water

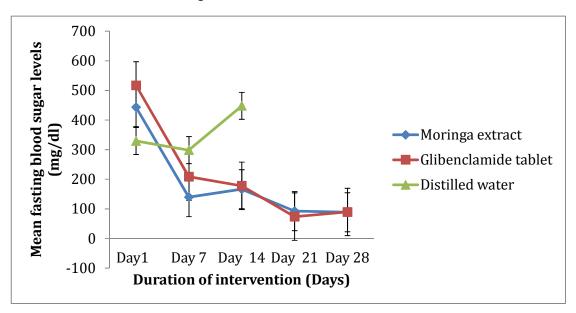


Fig. 2. Mean fasting blood sugar against days during intervention for *M. oleifera* aqueous extract glibenclamide tablet and distilled water

The mean body weight of the diabetic rats that received M. oleifera aqueous extract and glibenclamide increased slightly, although the increase was not significant. Glibenclamide is taken in dose of 5 mg daily and is clinically effective in lowering blood glucose and cause weight gain. Most type 2 diabetic patients experience weight loss if diabetes is not well controlled. Normal weight is an indicator of good clinical diabetic control. Although there is limited documentation of controlled studies that confirm M. oleifera leaves to cause weight loss, studies in human have shown that it reduces weight through its inhibition of α -amylase enzyme [26, 42]. However animals studies have shown increase in body weight when diabetic rats are treated with M. oleifera aqueous extract [21].

The leaves of *M. oleifera* are used in Ugandan rural communities to treat diabetes mellitus [16]. However, there is currently no recommended standard method of preparation and use in the Previous communities. preparations have involved maceration of fresh leaves of M. oleifera or powder in different quantities of water. This scenario encouraged us to carry out an experiment to compare the hyperglycemic control of M. oleifera leaves aqueous extract and Glibenclamide tablets in alloxan monohydrate induced diabetic rats. The results show that the aqueous leaf extract of Moringa oleifera compares well with the oral hypoglycemic drug glibenclamide in causing hypoglycemia in alloxan monohydrate induced diabetic rats

The study results agreed with those from earlier studies around the world which found out that M. oleifera leaves extracts reduced blood sugar in laboratory animals or has a hypoglycemic effect [21-33]. The fact that the quantity and quality of phytochemicals (non-nutritive secondary metabolites), which have medicinal principles, greatly depend on the soil, has encouraged many researchers from different regions and countries to explore the hypoglycemic effects of M. oleifera extracts grown locally in diabetic rat models [21-33]. Tannins. steroids and flavonoids. triterpenoids. saponins. anthraguinones, alkaloids and reducing sugars are phytochemicals that may have hypoglycemic effect also identified in M. oleifera extract [16, 34]. Each of thee phytocemicals or in combination may act together to reduce blood sugar in vivo.

It is believed that the hypoglycemic effect of *M. oleifera* is probably due to presence of phytochemicals that have anti-inflammatory

[31,32,33], antioxidant [25,26,27], and immunemodulatory [28] effects. Intake of flavonoids has been shown to protect against chronic diseases associated with oxidative stress, including cardiovascular disease, cancer and M. oleifera leaves are a good source of flavonoids [43]. Phenolic acids and flavonoids affect glucose homeostasis by influencing β-cell mass and function, plus increasing insulin sensitivity in peripheral tissues, which are good for diabetes prevention and management [44]. These compounds have also been shown to benefit patients with other chronic conditions such as. hypercholesterolemia, high blood pressure, nonalcoholic liver disease, and cancer [45]. Condensed tannin extracts showed promising antidiabetic effects with potential α -amylase and α -glucosidase inhibition activities [46] and the tannin-rich extract from plant material could be an interesting candidate for the treatment of several health disorders associated with oxidative stress such as hepatocellular injury and diabetes [47]. The hypoglycemic activity of M. oleifera leaves could also be due to presence of terpenoids, which appears to be involved in the stimulation of β cells and the subsequent secretion of preformed insulin [48]. Quercetin is found in dried M. Oleifera leaves, at high concentrations, as guercetin-3-O-β-d-glucoside (iso-quercetin or isotrifolin). Quercetin is a strong antioxidant, with multiple therapeutic properties including hypoglycemia [49]. It can protect insulin-producing pancreatic β cells from Streptozotocin (STZ) induced oxidative stress and apoptosis in rats [50]. In studies where diabetes is induced in rats doses as low as 250 mg/kg of M. oleifera leaves extract have caused hypoglycemia [33].

Despite the wealth of knowledge on the hypoglycemic effect of *M. oleifera* leaves in animal models, there is yet no drug developed from the plant phytochemicals to manage diabetes.

5. CONCLUSION

From this study we can conclude that *M. oleifera* leaves aqueous extract has similar pattern to glibenclamide tablet in causing hypoglycemia to alloxan monohydrate induced diabetic rats.

6. RECOMMENDATION

More studies are needed to develop hypoglycemic drugs from *M. oleifera* leaves in a

bid to effectively, safely and cheaply treat diabetes mellitus. Clinical trials in normal human volunteers to determine the safety of *M. oleifera* leaves extract.

CONSENT

It is not applicable.

ETHICAL APPROVAL

Ethical approval was obtained from the Institution Review Board (IRB) of the Makerere University College of Health Sciences, School of Biomedical Sciences, SBS-HDREC 565. The animals were treated humanely according to international standard OECD guidelines (2001).

ACKNOWLEDGEMENTS

Authors are grateful to The Directorate of Research and Graduate Training, CARNEGIE NERLP PROGRAMME, Makerere University for the financial support.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Global report on diabetes. World Health Organization, Geneva; 2016.
- 2. Uganda National diabetic report, WHO; 2016.
- Pradhan AD, Manson JE, Rifai N, Buring JE, Ridker PM. C-reactive protein, interleukin 6, and risk of developing type 2 diabetes mellitus. JAMA. 2001;286:327– 334.
- Spranger J, Kroke A, Mohlig M, Hoffmann K, Bergmann MM, Ristow M, Boeing H. Pfeiffer AF. Inflammatory cytokines and the risk to develop type 2 diabetes: Results of the prospective population-based European prospective investigation into cancer and nutrition (EPIC)-potsdam study. Diabetes. 2003;52:812–817.
- Hu FB, Meigs JB, Li TY, Rifai N, Manson JE. Inflammatory markers and risk of developing type 2 diabetes in women. Diabetes. 2004;53:693–700.
- Brandão-Lima P, Barbosa de Carvalho G, Santos RKF, Santos BC. NL Dias-

Vasconcelos; VSRK Barra; F Barbosa; Pires LV. Nutrients. 2018;10(12):1948.

- Thornalley PJ, et al. Diabetologia. High prevalence of low plasma thiamine concentration in diabetes linked to a marker of vascular disease. 2007;50: 2164–2170.
- 8. Smith AD, et al. Plos One. 2010;51–10.
- 9. Ceriello A, et al. Meal-generated oxidative stress in type 2 diabetic patients. Diabetes Care. 1998;21(9):1529–1533
- Montero D, et al. Effect of antioxidant vitamin supplementation on endothelial function in type 2 diabetes mellitus: a systematic review and meta-analysis of randomized controlled trials. Obes Rev. 2014;15(2):107–116.
- 11. Gröber U, et al. Update 2013: From rickets prophylaxis to general preventive healthcare. Dermatoendocrinol. 2013;5: 331–347
- 12. Holick MF. Nat Rev Endocrinol. 2012;8: 388–390.
- Zipitis CS. Vitamin D supplementation in early childhood and risk of type 1 diabetes: a systematic review and meta-analysis. Akobeng A. K. Arch Dis Child. 2008;93: 512–517
- 14. Warjeet Singh L. Traditional medicinal plants of Manipur as anti-diabetics. J. Med. Plants Res. 2011;5:677–687.
- 15. Farooq F, Rai M, Tiwari A, et al. Medicinal properties of *Moringa oleifera*: An overview of promising healer. J Med Plants Res. 2012;6:4368–4374.
- Kasolo JN, Bimenya GS, Lonzi O, Ochieng J, Ogwal-Okeng JW. Journal of Medicinal Plants Research. 2010;4(9):753-757.
- 17. Jed W. *Moringa oleifera*: A review of the medical evidence for its nutritional, therapeutic, and prophylactic properties. Trees of life Journal. 2005;1.
- 18. Fahey J. *Moringa oleifera*: A review of the medical evidence for its nutritional, therapeutic, and prophylactic properties. Part 1. Trees of life Journal. 2005;1(5).
- 19. Anwar F, Latir S, Ashraf M, Gilan A. Phytother Res. 2007;21:17-25.
- Gopalakrishnan L, Doriya K, Kumar DS. Moringa oleifera: A review on nutritive importance and its medicinal application. Food Science and Human Wellness. 2016; 5:49–56.
- 21. EIL Amira Abd, EIB Badr El Said, Mahboub HD, Mabrouk AAE. Biochem. Cell Biol. 2014;92.

- 22. Mbikay M. Therapeutic potential of *Moringa oleifera* leaves in chronic hyperglycemia and dyslipidemia: A review. Front Pharmacol. 2012;3:24.
- 23. Irfan Tang T, Choi EJ, Han WC, Oh M, Kim J, Hwang JY, Park PJ, Moon SH, Kim YS, Kim EK. J Med Food. 2017;2(5):502-510
- 24. Manohar VS, Jayasree T, Kishore KK, Rupa LM, Dixit R, Chandrasekhar NJ. Chem. Pharmacol. Res. 2012;4:249–253.
- 25. Gupta R, Manas M, Vijay K, Baja J, Katariya P, Yadav S, Kamal R, Gupta RS. Journal of Diabetes. 2012;4:164–171.
- Atawodi SE, Atawodi JS, Idakwo GA, Pfundstein B, Haubner R, Wurtele G, Bartsch H, Owen RW. J Med Food. 2010; 13:710-6.
- Kaneto H, Kajimoto Y, Miyagawa J, Matsuoka T, Fujitani Y, Umayahara Y, Hanafusa T, Matsuzawa Y, Yamasaki Y, Hori M. Diabetes. 1999;48:2398–2406.
- Nfambi J, Bbosa GS, Sembajwe LF, Gakunga J, Kasolo JN. J Basic Clin Physiol Pharmacol. 2015;6(6):603-11.
- 29. Jaiswa D, Kumar P, Kumar RA, Shikha M, Geeta W. Effect of *Moringa oleifera* Lam. leaves aqueous extract therapy on hyperglycemic rats. Journal of Ethnopharmacology. 2009;123(3):392-396.
- Divi SM, Bellamkonda R, Dasireddy SK. Asian J. Pharm. Clin. Res. 2012;5:67– 72.
- Sulaiman MR, Zakaria ZA, Bujarimin AS, Somchit MN, Israf DA, Moin S. Pharmaceutical Biology. 2009;838-845.
- 32. Coppin JP, Xu Y, Chen H, et al. Determination of flavonoids by LC/MS and anti-inflammatory activity in *Moringa oleifera.* J Funct Foods. 2013;5:1892– 1899.
- Omodanisi Aboua YG, Oguntibeju OO. Assessment of the anti-hyperglycaemic, anti-inflammatory and antioxidant activities of the methanol extract of *Moringa oleifera* in diabetes-induced nephrotoxic male wistar rats. Molecules. 2017;22:43.
- Bamishaiye EI, Olayemi FF, Awagu EF, Bamshaiye OM. Advance Journal of Food Science and Technology. 2011;3(4):233-237.
- Stohs SJ, Hartman MJ. Review of the safety and efficacy of *Moringa oleifera*. Phytother Res PTR. 2015;29:796–804.

- Kasolo JN, Bimenya GS, Okwi AL, Othieno EM, Ogwal-Okeng JW. Afri. J. Anim. Biomed. Sci. 2012;7(1):40-44.
- Adedapo AA, Mogbijuri OM, Emikpe BO. Journal of Medicinal Plants Research. 2009;3:586-591.
- Kasolo JN, Bimenya GS, Ojok L, Ogwalokeng JW. Sub-acute toxicity evaluation of *Moringa oleifera* leaves aqueous and ethanol extracts in Swiss Albino rats. International Journal of Medicinal Plant Research. 2012;1(6):075-081.
- Trivedi NA, Mazumdar B, Bhatt JD, Hemavathi KG. Indian Journal of Pharmacology. 2004;36(6):373-376.
- Harborne J. Methods of plant analysis Phytochemical methods. Springer. 1984;1-36.
- 41. Ciule I. Practical manuals on the industrial utilization of medicinal and aromatic plants, Romania, University of Bucharest; 1964.
- 42. Kashinath Dixit DM, Dinesh V, Kamath MD, Krishnaraju V, Alluri MS, Barbara A, Davis. Diabetes, obesity and metabolism. Journal of Pharmacology and Therapeutic. 2018;20:2633-2641.
- 43. Pandey KB, Rizvi SI. Plant polyphenols as dietary antioxidants in human health and disease. Oxid Med Cell Longev. 2009;2(5): 270-8.
- 44. Oboh G, Agunloye OM, Adefegha SA, Akinyemi AJ, Ademiluyi AO. J Basic Clin Physiol Pharmacol. 2015;26(2):165-70.
- 45. Vergara-Jimenez M, Almatrafi MM, Fernandez ML. Bioactive components in *Moringa oleifera* leaves protect against chronic disease. Antioxidants (Basel). 2017;6(4):91.
- Kunyanga CN, Imungi JK, Okoth M, Momanyi C, Biesalski HK, Vadivel V. Antioxidant and antidiabetic properties of condensed tannins in acetonic extract of selected raw and processed indigenous food ingredients from Kenya. J Food Sci. 2011;76(4):C560-7.
- Sobeh M, Mahmoud MF, Abdelfattah MAO, El-Beshbishy HA, El-Shazly AM, Wink M. Hepatoprotective and hypoglycemic effects of a tannin rich extract from *Ximenia americana* var. caffra root. Phytomedicine. 2017;15(33):36-42.
- 48. Tende Ezekiel I, Dikko AAU. Effect of ethanolic leaves extract of *Moringa oleifera* on blood glucose levels of streptozocininduced diabetics and normoglycemic

Wistar rats. British J Pharmacol Toxicol. 2011;2:1-4.

- 49. Bischoff SC. Efficacy of macronutrients on targeting obesity and beyond. Curr Opin Clin Nutr Metab Care. 2008;11(6):733-40.
- Coskun O, Kanter M, Korkmaz A, Oter S. Quercetin, a flavonoid antioxidant, prevents and protects streptozotocininduced oxidative stress and β-cell damage in rat pancreas. Pharmacol Res. 2005;51(2):117-23.

© 2019 Kasolo et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: http://www.sdiarticle3.com/review-history/48844